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(54) T7 DNA polymerase.

(57) This invention relates to T7-type DNA polymerases and methods for using them including a method for determining the nucleotide base sequence of a DNA molecule, comprising annealing said DNA molecule with a primer molecule able to hybridize to said DNA molecule; incubating separate portions of the annealed mixture in at least four vessels with four different deoxynucleoside triphosphates, a processive DNA polymerase, therein said polymerase remains bound to said DNA molecule for at least 500 bases before dissociating in an environmental condition normally used in the extension reaction of a DNA sequencing reaction, said polymeras having less than 500 units of exonuclease activity per mg of said polymerase, and one of four DNA synthesis terminating agents which terminate DNA synthesis at a specific nucleotide base. The agent terminates at a different specific nucleotide base in each of the four vessels. The DNA products of the incubating reaction are separated according to their side so that at least part of the nucleotide base sequence of the DNA molecule can be determined.

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T7 DNA POLYMERASE

This invention relates to DNA polymerases suitable for DNA sequencing and in particular relates to a purified modified gene encoding a modified DNA polymerase.

DNA sequencing involves the generation of four populations of single stranded DNA fragments having one defined terminus and one variable terminus. The variable terminus always terminates at a specific given nucleotide base (either guanine (G), adenine (A), thymine (T), or cytosine (C)). The four different sets of fragments are each separated on the basis of their length, on a high resolution polyacrylamide gel; each band on the gel corresponds colinearly to a specific nucleotide in the DNA sequence, thus identifying the positions in the sequence of the given nucleotide base.

Generally there are two methods of DNA sequencing. One method (Maxam and Gilbert sequencing) involves the chemical degradation of isolated DNA fragments, each labelled with a single radiolabel at its defined terminus, each reaction yielding a limited cleavage specifically at one ore more of the four bases (G, A, T or C). The other method (dideoxy sequencing) involves the enzymatic synthesis of a DNA strand. Four separate syntheses are run, each reaction being caused to terminate at a specific base (G, A, T or C) via incorporation of the appropriate chain terminating dideoxynucleotide. The latter method is preferred since the DNA fragments are uniformly labelled (instead of end labelled) and thus the larger DNA fragments contain increasingly more radioactivity. Further, ^{35}S-labelled nucleotides can be used in place of ^{32}P-labelled nucleotides, resulting in sharper definition; and the reaction products are simple to interpret since each lane corresponds only to either G, A, T or C. The enzyme used for most dideoxy sequencing is the Escherichia coli DNA-polymerase I large fragment ("Klenow"). Another polymerase used is AMV reverse transcriptase.

Summary of the Invention

In one aspect the invention features a method for determining the nucleotide base sequence of a DNA molecule, comprising annealing the DNA molecule with a primer molecule able to hybridize to the DNA molecule; incubating separate portions of the annealed mixture in at least four vessels with four different deoxynucleoside triphosphates, a processive DNA polymerase wherein the polymerase remains bound to a DNA molecule for at least 500 bases before dissociating in an environmental condition normally used in the extension reaction of a DNA sequencing reaction, the polymerase having less than 500 units of exonuclease activity per mg of polymerase, and one of four DNA synthesis terminating agents which terminate DNA synthesis at a specific nucleotide base. The agent terminates at a different specific nucleotide base in each of the four vessels. The DNA products of the incubating reaction are separated according to their size so that at least a part of the nucleotide base sequence of the DNA molecule can be determined.

In preferred embodiments the polymerase remains bound to the DNA molecule for at least 1000 bases before dissociating; the polymerase is substantially the same as one in cells infected with a T7-type phage (i.e., phage in which the DNA polymerase requires host thioredoxin as a subunit; for example, the T7-type phage is T7, T3, ϕI, ϕII, H, W31, gh-1, Y, A1122, or SP6, Studier, 95 Virology 70, 1979); the polymerase is non-discriminating for dideoxy nucleotide analogs; the polymerase is modified to have less than 50 units of exonuclease activity per mg of polymerase, more preferably less than 1 unit, even more preferably less than 0.1 unit, and most preferably has no detectable exonuclease activity; the polymerase is able to utilize primers of as short as 10 bases or preferably as short as 4 bases; the primer comprises four to forty nucleotide bases, and is single stranded DNA or RNA; the annealing step comprises heating the DNA molecule and the primer to above 65° C, preferably from 65° C to 100° C. and allowing the heated mixture to cool to below 65° C, preferably to 0° C to 30° C; the incubating step comprises a pulse and a chase step, wherein the pulse step comprises mixing the annealed mixture with all four different deoxynucleoside triphosphates and a processive DNA polymerase wherein at least one of the deoxynucleoside triphosphates is labelled; most preferably the pulse step performed under conditions in which the polymerase does not exhibit its processivity and is for 30 seconds to 20 minutes at 0° C to 20° C or where at least one of the nucleotid triphosphates is limiting; and the chase step comprises adding one of the chain terminating agents to four separate aliquots of the mixture after the pulse step; preferably the chase step is for 1 to 60 minutes at 30° C to 50° C; the terminating agent is a dideoxynucleotide, or a limiting level of one deoxynucleoside triphosphat ; one of the four deoxynucleotides is dITP or deazaguanosine; labelled primers are us d so that no pulse step is required, preferably the label is radioactive or fluorescent; and the

polymerase is unable to exhibit its processivity in a second nvironmental condition normally used in the pulse reaction of a DNA sequencing reaction.

In other aspects the invention features a) a method for producing blunt ended double-stranded DNA molecules from a linear DNA molecule having no 3′ protruding termini, using a processive DNA polymerase free from exonuclease activity; b) a method of amplification of a DNA sequence comprising annealing a first and second primer to opposite strands of a double stranded DNA sequence and incubating the annealed mixture with a processive DNA polymerase having less than 500 units of exonuclease activity per mg of polymerase, preferably less than 1 unit, wherein the first and second primers anneal to opposite strands of the DNA sequence; in preferred embodiments the primers have their 3′ ends directed toward each other; and the method further comprises, after the incubation step, denaturing the resulting DNA, annealing th first and second primers to the resulting DNA and incubating the annealed mixture with the polymerase; preferably the cycle of denaturing, annealing and incubating is repeated from 10 to 40 times; c) a method for in vitro mutagenesis of cloned DNA fragments, comprising providing a cloned fragment and synthesizing a DNA strand using a processive DNA polymerase having less than 1 unit of exonuclease activity per mg of polymerase; d) a method of producing active T7-type DNA polymerase from cloned DNA fragments under the control of non-leaky promoters (see below) in the same cell comprising inducing expression of the genes only when the cells are in logarithmic growth phase, or stationary phase, and isolating the polymerase from the cell; preferably the cloned fragments are under the control of a promoter requiring T7 RNA polymerase for expression; e) a gene encoding a T7-type DNA polymerase, the gene being genetically modified to reduce the activity of naturally occurring exonuclease activity; most preferably a histidine (His) residue is modified, even more preferably His-123 of gene 5; f) the product of the gene encoding genetically modified polymerase; g) a method of purifying T7 DNA polymerase from cells comprising a vector from which the polymerase is expressed, comprising the steps of lysing the cells, and passing the polymerase over an ion-exchange column, over a DE52 DEAE column, a phosphocellulose column, and a hydroxyapatite column; preferably prior to the passing step the method comprises precipitating the polymerase with ammonium sulfate; the method further comprises the step of passing the polymerase over a Sephadex DEAE A50 column; and the ion-exchange column is a DE52 DEAE column; h) a method of inactivating exonuclease activity in a DNA polymerase solution comprising incubating the solution in a vessel containing oxygen, a reducing agent and a transition metal; i) a kit for DNA sequencing, comprising a processive DNA polymerase, defined as above, having less than 500 units of exonucleas activity per mg of polymerase, wherein the polymerase is able to exhibit its processivity in a first environmental condition, and preferably unable to exhibit its processivity in a second environmental condition, and a reagent necessary for the sequencing, selected from a chain terminating agent, and dITP: j) a method for labelling the 3′ end of a DNA fragment comprising incubating the DNA fragment with a processive DNA polymerase having less than 500 units of exonuclease activity per mg of polymerase, and a labelled deoxynucleotide; k) a method for in vitro mutagenesis of a cloned DNA fragment comprising providing a primer and a template, the primer and the template having a specific mismatched base, and extending the primer with a processive DNA polymerase; and 1) a method for in vitro mutagenesis of a cloned DNA fragment comprising providing the cloned fragment and synthesizing a DNA strand using a processive DNA polymerase, having less than 50 units of exonuclease activity, under conditions which cause misincorporation of a nucleotide base.

This invention provides a DNA polymerase which is processive, non-discriminating, and can utilize short primers. Further, the polymerase has no associated exonuclease activity. These are ideal properties for the above described methods, and in particular for DNA sequencing reactions, since the background level of radioactivity in the polyacylamide gels is negligible, there are few or no artifactual bands, and the bands are sharp -- making the DNA sequence easy to read. Further, such a polymerase allows novel methods of sequencing long DNA fragments, as is described in detail below.

Other features and advantages of the invention will be apparent from the following description of the preferred embodiments thereof and from the claims.

D scription of the Pr f rr d Embodiments

The drawings will first briefly be described.

Drawings

Figs. 1-3 are diagrammatic representations of the vectors pTrx-2, mGP1-1, and pGP5-5 respectively;

Fig. 4 is a graphical repr sentation of th selective oxidation of T7 DNA polym rase;

Fig. 5 is a graphical representation of the ability of modified T7 polymerase to synthesiz DNA in the presence of etheno-dATP; and

Fig. 6 is a diagrammatic representation of the enzymatic amplification of genomic DNA using modified T7 DNA polymerase.

Fig. 7, 8 and 9 are the nucleotide sequences of pTrx-2, a part of pGP5-5 and mGP1-2 respectively.

Fig. 10 is a diagrammatic representation of pGP5-6.

DNA Polymerase

In general the DNA polymerase of this invention is processive, has no associated exonuclease activity, does not discriminate against nucleotide analog incorporation, and can utilize small oligonucleotides (such as tetramers, hexamers and octamers) as specific primers. These properties will now be discussed in detail.

Processivity

By processivity is meant that the DNA polymerase is able to continuously incorporate many nucleotides using the same primer-template without dissociating from the template, under conditions normally used for DNA sequencing extension reactions. The degree of processivity varies with different polymerases: some incorporate only a few bases before dissociating (e.g. Klenow (about 15 bases), T4 DNA polymerase (about 10 bases), T5 DNA polymerase (about 180 bases) and reverse transcriptase (about 200 bases) (Das et al. J. Biol. Chem. 254:1227 1979; Bambara et al., J. Biol. Chem 253:413, 1978) while others, such as those of the present invention, will remain bound for at least 500 bases and preferably at least 1,000 bases under suitable environmental conditions. Such environmental conditions include having adequate supplies of all four deoxynucleoside triphosphates and an incubation temperature from 10° C-50° C. Processivity is greatly enhanced in the presence of E. coli single stranded binding (ssb), protein.

With processive enzymes termination of a sequencing reaction will occur only at those bases which have incorporated a chain terminating agent, such as a dideoxynucleotide. If the DNA polymerase is non-processive, then artifactual bands will arise during sequencing reactions, at positions corresponding to the nucleotide where the polymerase dissociated. Frequent dissociation creates a background of bands at incorrect positions and obscures the true DNA sequence. This problem is partially corrected by incubating the reaction mixture for a long time (30-60 min) with a high concentration of substrates, which "chase" the artifactual bands up to a high molecular weight at the top of the gel, away from the region where the DNA sequence is read. This is not an ideal solution since a non-processive DNA polymerase has a high probability of dissociating from the template at regions of compact secondary structure, or hairpins. Reinitiation of primer elongation at these sites is inefficient and the usual result is the formation of bands at the same position for all four nucleotides, thus obscuring the DNA sequence.

Analog discrimation

The DNA polymerases of this invention do not discriminate significantly between dideoxy-nucleotide analogs and normal nucleotides. That is, the chance of incorporation of an analog is approximately the same as that of a normal nucleotide or at least incorporates the analog with at least 1/10 the efficiency that of a normal analog. The polymerases of this invention also do not discriminate significantly against some other analogs. This is important since, in addition to the four normal deoxynucleoside triphosphates (dGTP, dATP, dTTP and dCTP), sequencing reactions require the incorporation of other types of nucleotide derivatives such as: radioactively-or fluorescently-labelled nucleoside triphosphates, usually for labeling the synthesized strands with ^{35}S, ^{32}P, or other chemical agents. When a DNA polymerase does not discriminate against analogs the same probability will exist for the incorporation of an analog as for a normal nucleotide. For labelled nucleoside triphosphates this is important in order to efficiently label the synthesized DNA strands using a minimum of radioactivity. Further, lower levels of analogs are required with such enzymes, making the sequencing reaction cheaper than with a discriminating enzyme.

Discriminating polymerases show a different xtent of discrimination when they are polymerizing in a processive mode versus when stalled, struggling to synthesize through a secondary structure impediment. At such impedim nts there will be a variability in the intensity of different radioactiv bands on the g l,

which may obscure the sequence.

Exonuclease Activity

The DNA polymerase of the invention has less than 50%, preferably less than 1%, and most preferably less than 0.1%, of the normal or naturally associated level of exonuclease activity (amount of activity per polymerase molecule). By normal or naturally associated level is meant the exonuclease activity of unmodified T7-type polymerase. Normally the associated activity is about 5,000 units of exonuclease activity per mg of polymerase, measured as described below by a modification of the procedure of Chase et al. (249 J. Biol. Chem. 4545, 1974). Exonucleases increase the fidelity of DNA synthesis by excising any newly synthesized bases which are incorrectly basepaired to the template. Such associated exonuclease activities are detrimental to the quality of DNA sequencing reactions. They raise the minimal required concentration of nucleotide precursors which must be added to the reaction since, when the nucleotide concentration falls, the polymerase activity slows to a rate comparable with the exonuclease activity, resulting in no net DNA synthesis, or even degradation of the synthesized DNA.

More importantly, associated exonuclease activity will cause a DNA polymerase to idle at regions in the template with secondary structure impediments. When a polymerase approaches such a structure its rate of synthesis decreases as it struggles to pass. An associated exonuclease will excise the newly synthesized DNA when the polymerase stalls. As a consequence numerous cycles of synthesis and excision will occur. This may result in the polymerase eventually synthesizing past the hairpin (with no detriment to the quality of the sequencing reaction); or the polymerase may dissociate from the synthesized strand (resulting in an artifactual band at the same position in all four sequencing reactions); or, a chain terminating agent may be incorporated at a high frequency and produce a wide variability in the intensity of different fragments in a sequencing gel. This happens because the frequency of incorporation of a chain terminating agent at any given site increases with the number of opportunities the polymerase has to incorporate the chain terminating nucleotide, and so the DNA polymerase will incorporate a chain-terminating agent at a much higher frequency at sites of idling than at other sites.

An ideal sequencing reaction will produce bands of uniform intensity throughout the gel. This is essential for obtaining the optimal exposure of the X-ray film for every radioactive fragment. If there is variable intensity of radioactive bands, then fainter bands have a chance of going undetected. To obtain uniform radioactive intensity of all fragments, the DNA polymerase should spend the same interval of time at each position on the DNA, showing no preference for either the additon or removal of nucleotides at any given site. This occurs if the DNA polymerase lacks any associated exonuclease, so that it will have only one opportunity to incorporate a chain terminating nucleotide at each position along the template.

Short primers

The DNA polymerase of the invention is able to utilize primers of 10 bases or less, as well as longer ones, most preferably of 4-20 bases. The ability to utilize short primers offers a number of important advantages to DNA sequencing. The shorter primers are cheaper to buy and easier to synthesize than the usual 15-20-mer primers. They also anneal faster to complementary sites on a DNA template, thus making the sequencing reaction faster. Further, the ability to utilize small (e.g., six or seven base) oligonucleotide primers for DNA sequencing permits strategies not otherwise possible for sequencing long DNA fragments. For example, a kit containing 80 random hexamers could be generated, none of which are complementary to any sites in the cloning vector. Statistically, one of the 80 hexamer sequences will occur an average of every 50 bases along the DNA fragment to be sequenced. The determination of a sequence of 3000 bases would require only five sequencing cycles. First, a "universal" primer (e.g., New England Biolabs #1211, sequence 5′ GTAAAACGACGGCCAGT 3′) would be used to sequence about 600 bases at one end of the insert. Using the results from this sequencing reaction, a new primer would be picked from the kit homologous to a region near the end of the determin d sequence. In the second cycle, the sequence of the next 600 bases would be d termined using this primer. Repetition of this process five times would det rmine the complete sequence of the 3000 bases, without necessitating any subcloning, and without the chemical synthesis of any new oligonucl otide primers. The use of such short prim rs may be enhanced by including gene 2.5 and 4 protein of T7 in the sequencing reaction.

DNA polymerases of this invention, (i.e., having the above properties) includ modified T7-type polymeras s. That is the DNA polymerase requires host thioredoxin as a sub-unit, and they are substan-

tially identical to a modified T7 DNA polymerase or to equivalent enzymes isolated from r lated phage, such as T3, ϕI, ϕII, H, W31, gh-1, Y, A1122 and SP6. Each of these enzymes can be modified to have prop rties similar to those of the modified T7 enzyme. It is possible to isolate the enzyme from phage infected cells directly, but preferably the enzyme is isolated from cells which overproduce it. By substantially identical is meant that the enzyme may have amino acid substitutions which do not affect the overall properties of the enzyme. One example of a particularly desirable amino acid substitution is one in which the natural enzyme is modified to remove any exonuclease activity. This modification may be performed at the genetic or chemical level (see below).

Cloning T7 polymerase

As an example of the invention we shall describe the cloning, overproduction, purification, modification and use of T7 DNA polymerase. This processive enzyme consists of two polypeptides tightly complexed in a one to one stoichiometry. One is the phage T7-encoded gene 5 protein of 84,000 daltons (Modrich et al. 150 J. Biol. Chem. 5515, 1975), the other is the E. coli encoded thioredoxin, of 12,000 daltons (Tabor et al., J. Biol, Chem. 262:16, 216, 1987). The thioredoxin is an accessory protein and attaches the gene 5 protein (the non-processive actual DNA polymerase) to the primer template. The natural DNA polymerase has a very active 3′ to 5′ exonuclease associated with it. This activity makes the polymerase useless for DNA sequencing and must be inactivated or modified before the polymerase can be used. This is readily performed, as described below, either chemically, by local oxidation of the exonuclease domain, or genetically, by modifying the coding region of the polymerase gene encoding this activity.

pTrx-2

In order to clone the trxA (thioredoxin) gene of E. coli wild type E. coli DNA was partially cleaved with Sau3A and the fragments ligated to BamHI-cleaved T7 DNA isolated from strain T7 ST9 (Tabor et al., in Thioredoxin and Glutaredoxin Systems: Structure and Function (Holmgren et al., eds) pp. 285-300, Raven Press, NY; and Tabor et al., supra). The ligated DNA was transfected into E. coli trxA⁻ cells, the mixture plated onto trxA⁻ cells, and the resulting T7 plaques picked. Since T7 cannot grow without an active E. coli trxA gene only those phages containing the trxA gene could form plaques. The cloned trxA genes were located on a 470 base pair HincII fragment.

In order to overproduce thioreodoxin a plasmid, pTrx-2, was as constructed. Briefly, the 470 base pair HincII fragment containing the trxA gene was isolated by standard procedure (Maniatis et al., Cloning: A Laboratory Manual, Cold Spring Harbor Labs., Cold Spring Harbor, N.Y.), and ligated to a derivative of pBR322 containing a Ptac promoter (ptac-12, Amann et al., 25 Gene 167, 1983). Referring to Fig. 2, ptac-12, containing β-lactamase and Col EI origin, was cut with PvuII, to yield a fragment of 2290 bp, which was then ligated to two tandem copies of trxA (HincII fragment) using commercially available linkers (SmaI-BamHI polylinker), to form pTrx-2. The complete nucleotide sequence of pTrx-2 is shown in Figure 7. Thioredoxin production is now under the control of the tac promoter, and thus can be specifically induced, e.g. by IPTG (isopropyl β-D-thiogalactoside).

pGP5-5 and mGP1-2

Some gene products of T7 are lethal when expressed in E. coli. An expression system was developed to facilitate cloning and expression of, lethal genes, based on the inducible expression of T7 RNA polymerase. Gene 5 protein is lethal in some E. coli strains and an example of such a system is described by Tabor et al. 82 Proc. Nat. Acad. Sci. 1074 (1985) where T7 gene 5 was placed under the control of the ϕ10 promoter, and is only expressed when T7 RNA polymerase is present in the cell.

Briefly, pGP5-5 (Fig. 3) was constructed by standard procedures using synthetic BamHI linkers to join T7 fragm nt from 14306 (Nd I) to 16869 (AhaIII), containing gene 5, to the 560 bp fragment of T7 from 5667 (HincII) to 6166 (Fnu4H1) containing both the ϕ1.1A and ϕ1.1B promoters, which are recognized by T7 RNA polymerase, and the 3kb BamHI-HincII fragment of pACYC177 (Chang et al., 134 J. Bacteriol. 1141, 1978). The nucl otide sequence of the T7 inserts and linkers in shown in Fig. 8. In this plasmid gene 5 is only expr ssed when T7 RNA polymerase is provided in the cell.

Referring to Fig. 3, T7 RNA polymerase is provided on phage vector mGP1-2. This is similar to pGP1-2

(Tabor et al., id.) xc pt that the fragment of T7 from 3133 (HaeIII) to 5840 (HinfI), containing T7 RNA polymerase was ligated, using linkers (BglII and SalI respectively), to BamHI-SalI cut M13 mp8, placing th polymerase gene under control of the lac promoter. The complete nucleotid sequenc of mGP1-2 is shown in Fig. 9.

Since pGP5-5 and pTrx-2 have different origins of replication (respectively a P15A and a ColE1 origin) they can be tranformed into one cell simultaneously. pTrx-2 expresses large quantities of thioredoxin in the presence of IPTG. mGP1-2 can coexist in the same cell as these two plasmids and be used to regulate expression of T7-DNA polymerase from pGP5-5, simply by causing production of T7-RNA polymerase by inducing the lac promoter with, e.g., IPTG.

Overproduction of T7 DNA polymerase

There are several potential strategies for overproducing and reconstituting the two gene products of trxA and gene 5. The same cell strains and plasmids can be utilized for all the strategies. In the preferred strategy the two genes are co-overexpressed in the same cell. (This is because gene 5 is susceptible to proteases until thioredoxin is bound to it.) As described in detail below, one procedure is to place the two genes separately on each of two compatible plasmids in the same cell. Alternatively, the two genes could be placed in tandem on the same plasmid. It is important that the T7-gene 5 is placed under the control of a non-leaky inducible promoter, such as ϕ1.1A, ϕ1.1B and ϕ10 of T7, as the synthesis of even small quantities of the two polypeptides together is toxic in most E. coli cells. By non-leaky is meant that less than 500 molecules of the gene product are produced, per cell generation time, from the gene when the promoter, controlling the gene's expression, is not activated. Preferably the T7 RNA polymerase expression system is used although other expression systems which utilize inducible promoters could also be used. A leaky promoter, e.g., plac, allows more than 500 molecules of protein to be synthesized, even when not induced, thus cells containing lethal genes under the control of such a promoter grow poorly and are not suitable in this invention. It is of course possible to produce these products in cells where they are not lethal, for example, the plac promoter is suitable in such cells.

In a second strategy each gene can be cloned and overexpressed separately. Using this strategy, the cells containing the individually overproduced polypeptides are combined prior to preparing the extracts, at which point the two polypeptides form an active T7 DNA polymerase.

Example 1: Production of T7 DNA polymerase

E. coli strain 71.18 (Messing et al., Proc. Nat. Acad. Sci. 74:3642, 1977) is used for preparing stocks of mGP1-2. 71.18 is stored in 50% glycerol at -80° C. and is streaked on a standard minimal media agar plat . A single colony is grown overnight in 25 ml standard M9 media at 37° C, and a single plaque of mGP1-2 is obtained by titering the stock using freshly prepared 71.18 cells. The plaque is used to inoculate 10 ml 2X LB (2% Bacto-Tryptone, 1% yeast extract, 0.5% NaCl, 8mM NaOH) containing JM103 grown to an $A_{590} = 0.5$. This culture will provide the phage stock for preparing a large culture of mGP1-2. After 3-12 hours, the 10 ml culture is centrifuged, and the supernatant used to infect the large (2L) culture. For the large culture, 4 X 500 ml 2X LB is inoculated with 4 X 5 ml 71.18 cells grown in M9, and is shaken at 37° C. When the large culture of cells has grown to an $A_{590} = 1.0$ (approximately three hours), they are inoculated with 10 ml of supernatant containing the starter lysate of mGP1-2. The infected cells are then grown overnight at 37° C. The next day, the cells are removed by centrifugation, and the supernatant is ready to use for induction of K38/pGP5-5/pTrx-2 (see below). The supernatant can be stored at 4° C for approximately six months, at a titer ~5 X 10^{11} ϕ/ml. At this titer, 1 L of phage will infect 12 liters of cells at an $A_{590} = 5$ with a multiplicity of infection of 15. If the titer is low, the mGP1-2 phage can be concentrated from the supernatant by dissolving NaCl (60 gm/liter) and PEG-6000 (65 gm/liter) in the supernatant, allowing the mixture to settle at 0° C for 1-72 hours, and then centrifuging (7000 rpm for 20 min). The precipitate, which contains the mGP1-2 phage, is resuspended in approximately 1/20th of the original volume of M9 media.

K38/pGP5-5/pTrx-2 is the E. coli strain (genotype HfrC (λ)) containing the two compatible plasmids pGP5-5 and pTrx-2. pGP5-5 plasmid has a P15A origin of replication and expresses th kanamycin (Km) resistance gene. pTrx-2 has a ColEI origin of r plication and expresses the ampicillin (Ap) r sistance gene. The plasmids ar introduced into K38 by standard procedures, selecting Km^R and Ap^R respectively. The cells K38/pGP5-5/pTrx-2 are stored in 50% glycerol at -80° C. Prior to use th y are streaked on a plate containing 50μg/ml ampicillin and kanamycin, grown at 37° C ov rnight, and a single colony grown in 10 ml

LB media containing 50μg/ml ampicillin and kanamycin, at 37° C for 4-6 hours. The 10 ml cell culture is used to inoculate 500 ml of LB media containing 50μg/ml ampicillin and kanamycin and shaken at 37° C overnight. The following day, the 500 ml culture is used to inoculate 12 lit rs of 2X LB-KPO_4 m dia (2% Bacto-Tryptone, 1% yeast extract, 0.5% NaCl, 20 mM KPO_4, 0.2% dextrose, and 0.2% casamino acids, pH 7.4), and grown with aeration in a fermentor at 37° C. When the cells reach an $A_{590} = 5.0$ (i.e. logarithmic or stationary phase cells), they are infected with mGP1-2 at a multiplicity of infection of 10, and IPTG is added (final concentration 0.5mM). The IPTG induces production of thioredoxin and the T7 RNA polymerase in mGP1-2, and thence induces production of the cloned DNA polymerase. The cells are grown for an additional 2.5 hours with stirring and aeration, and then harvested. The cell pellet is resuspended in 1.5 L 10% sucrose/20 mM Tris-HCl, pH 8.0/25 mM EDTA and re-spun. Finally, the cell pellet is resuspended in 200 ml 10% sucrose/20 mM Tris-HCl, pH 8/1.0 mM EDTA, and frozen in liquid N_2. From 12 liters of induced cells 70 gm of cell paste are obtained containing approximately 700 mg gene 5 protein and 100 mg thioredoxin.

K38/pTrx-2 (K38 containing pTrx-2 alone) overproduces thioredoxin, and it is added as a "booster" to extracts of K38/pGP5-5/pTrx-2 to insure that thioredoxin is in excess over gene 5 protein at the outset of the purification. The K38/pTrx-2 cells are stored in 50% glycerol at -80° C. Prior to use they are streaked on a plate containing 50 μg/ml ampicillin, grown at 37° C for 24 hours, and a single colony grown at 37° C overnight in 25 ml LB media containing 50 μg/ml ampicillin. The 25 ml culture is used to inoculate 2 L of 2X LB media and shaken at 37° C. When the cells reach an $A_{590} = 3.0$, the ptac promoter, and thus thioredoxin production, is induced by the addition of IPTG (final concentration 0.5 mM). The cells are grown with shaking for an additional 12-16 hours at 37° C, harvested, resuspended in 600 ml 10% sucrose/20 mM Tris-HCl, pH 8.0/25 mM EDTA, and re-spun. Finally, the cells are resuspended in 40 ml 10% sucrose/20 mM Tris-HCl, pH 8/0.5 mM EDTA, and frozen in liquid N_2. From 2L of cells 16 gm of cell paste are obtained containing 150 mg of thioredoxin.

Assays for the polymerase involve the use of single-stranded calf thymus DNA (6mM) as a substrate. This is prepared immediately prior to use by denaturation of double-stranded calf thymus DNA with 50 mM NaOH at 20° C for 15 min., followed by neutralization with HCl. Any purified DNA can be used as a template for the polymerase assay, although preferably it will have a length greater than 1,000 bases.

The standard T7 DNA polymerase assay used is a modification of the procedure described by Grippo et al. (246 J. Biol. Chem. 6867, 1971). The standard reaction mix (200 μl final volume) contains 40 mM Tris/HCl pH 7.5, 10 mM $MgCl_2$, 5 mM dithiothreitol, 100 nmol alkali-denatured calf thymus DNA, 0.3 dGTP, dATP, dCTP and [^{3}H]dTTP (20 cpm/pm), 50 μg/ml BSA, and varying amounts of T7 DNA polymerase. Incubation is at 37° C (10° C-45° C) for 30 min (5 min-60 min). The reaction is stopped by the addition of 3 ml of cold (0° C) 1 N HCl-0.1 M pyrophosphate. Acid-insoluble radioactivity is determined by the procedure of Hinkle et al. (250 J. Biol. Chem. 5523, 1974). The DNA is precipitated on ice for 15 min (5 min-12 hr), then precipitated onto glass-fiber filters by filtration. The filters are washed five times with 4 ml of cold (0° C) 0.1M HCl-0.1M pyrophosphate, and twice with cold (0° C) 90% ethanol. After drying, the radioactivity on the filters is counted using a non-aqueous scintillation fluor.

One unit of polymerase activity catalyzes the incorporation of 10 nmol of total nucleotide into an acid-soluble form in 30 min at 37° C, under the conditions given above. Native T7 DNA polymerase and modified T7 DNA polymerase (see below) have the same specific polymerase activity ± 20%, which ranges between 5,000-20,000 units/mg for native and 5,000-50,000 units/mg for modified polymerase) depending upon the preparation, using the standard assay conditions stated above.

T7 DNA polymerase is purified from the above extracts by precipitation and chromatography techniques. An example of such a purification follows.

An extract of frozen cells (200 ml K38/pGP5-5/pTrx-2 and 40 ml K38/pTrx-2) are thawed at 0° C overnight. The cells are combined, and 5 ml of lysozyme (15 mg/ml) and 10 ml of NaCl (5M) are added. After 45 min at 0° C, the cells are placed in a 37° C water bath until their temperature reaches 20° C. The cells are then frozen in liquid N_2. An additional 50 ml of NaCl (5M) is added, and the cells are thawed in a 37° C water bath. After thawing, the cells are gently mixed at 0° C for 60 min. The lysate is centrifuged for one hr at 35,000 rpm in a Beckman 45Ti rotor. The supernatant (250 ml) is fraction I. It contains approximately 700 mg gene 5 protein and 250 mg of thioredoxin (a 2:1 ratio thioredoxin to gene 5 protein).

90 gm of ammonium sulphate is dissolved in fraction I (250 ml) and stirred for 60 min. The suspension is allowed to sit for 60 min, and the resulting precipitate collected by centrifugation at 8000 rpm for 60 min. The precipitate is redissolved in 300 ml of 20 mM Tris-HCl pH 7.5/5 mM 2-mercaptoethanol/0.1 mM EDTA/10% glycerol (Buffer A). This is fraction II.

A column of Whatman DE52 DEAE (12.6 cm^2 x 18 cm) is prepared and washed with Buffer A. Fraction II is dialyz d ov rnight against two changes of 1 L of Buffer A each until the conductivity of Fraction II has a

conductivity equal to that of Buffer A containing 100 mM NaCl. Dialyzed Fraction II is applied to th column at a flow rate of 100 ml/hr, and washed with 400 ml of Buffer A containing 100 NaCl. Proteins are eluted with a 3.5 L gradient from 100 to 400 mM NaCl in Buffer A at a flow rate of 60 ml/hr. Fractions containing T7 DNA polymerase, which elutes at 200 mM NaCl, are pooled. This is fraction III (190 ml).

A column of Whatman P11 phosphocellulose (12.6 cm^2 x 11 cm) is prepared and washed with 20 mM KPO_4 pH 7.4/5 mM 2-mercaptoethanol/0.1 mM EDTA/10 % glycerol (Buffer B). Fraction III is diluted 2-fold (380 ml) with Buffer B, then applied to the column at a flow rate of 60 ml/hr and washed with 200 ml of Buffer B containing 100mM KCl. Proteins are eluted with a 1.8 L gradient from 100 to 400 mM KCl in Buffer B at a flow rate of 60 ml/hr. Fractions containing T7 DNA polymerase which elutes at 300 KCl, are pooled. This is fraction IV (370 ml).

A column of DEAE-Sephadex A-50 (4.9 cm^2 x 15 cm) is prepared and washed with 20 mM Tris-HCl 7.0/0.1 mM dithiothreitol/0.1 mM EDTA/10% glycerol (Buffer C). Fraction IV is dialyzed against two changes of 1 L Buffer C to a final conductivity equal to that of Buffer C containing 100 mM NaCl. Dialyzed fraction IV is applied to the column at a flow rate of 40 ml/hr, and washed with 150 ml of Buffer C containing 100 mM NaCl. Proteins are eluted with a 1 L gradient from 100 to 300 mM NaCl in Buffer C at a flow rate of 40 ml/hr. Fractions containing T7 DNA polymerase, which elutes at 210 mM NaCl, are pooled. This is fraction V (120 ml).

A column of BioRad HTP hydroxylapatite (4.9 cm^2 x 15 cm) is prepared and washed with 20 mM KPO_4, pH 7.4/10 mM 2-mercaptoethanol/2 mM Na citrate/10% glycerol (Buffer D). Fraction V is dialyzed against two changes of 500 ml Buffer D each. Dialyzed fraction V is applied to the column at a flow rate of 30 ml/hr, and washed with 100 ml of Buffer D. Proteins are eluted with a 900 ml gradient from 0 to 180 mM KPO_4, pH 7.4 in Buffer D at a flow rate of 30 ml/hr. Fractions containing T7 DNA polymerases which elutes at 50 mM KPO_4, are pooled. This is fraction VI (130 ml). It contains 270 mg of homogeneous T7 DNA polymerase.

Fraction VI is dialyzed versus 20 mM KPO_4 pH 7.4/0.1 mM dithiothreitol/0.1 mM EDTA/50% glycerol. This is concentrated fraction VI (~65 ml, 4 mg/ml), and is stored at -20° C.

The isolated T7 polymerase has exonuclease activity associated with it. As stated above this must be inactivated. An example of inactivation by chemical modification follows.

Concentrated fraction VI is dialyzed overnight against 20 mM KPO_4 pH 7.4/0.1 mM dithiothreitol/10% glycerol to remove the EDTA present in the storage buffer. After dialysis, the concentration is adjusted to 2 mg/ml with 20 mM KPO_4 pH 7.4/0.1 mM dithiothreitol/10% glycerol, and 30 ml (2mg/ml) aliquots are placed in 50 ml polypropylene tubes. (At 2 mg/ml, the molar concentration of T7 DNA polymerase is 22 µM.)

Dithiothreitol (DTT) and ferrous ammonium sulfate ($Fe(NH_4)_2(SO_4)_26H_2O$) are prepared fresh immediately before use, and added to a 30 ml aliquot of T7 DNA polymerase, to concentrations of 5 mM DTT (0.6 ml of a 250 stock) and 20µM $Fe(NH_4)_2(SO_4)_26H_2O$ (0.6 ml of a 1 mM stock). During modification the molar concentrations of T7 DNA polymerase and iron are each approximately 20 µM, while DTT is in 250X molar excess.

The modification is carried out at 0° C under a saturated oxygen atmosphere as follows. The reaction mixture is placed on ice within a dessicator, the dessicator is purged of air by evacuation and subsequently filled with 100% oxygen. This cycle is repeated three times. The reaction can be performed in air (20% oxygen), but occurs at one third the rate.

The time course of loss of exonuclease activity is shown in Fig. 4. ^3H-labeled double-stranded DNA (6 cpm/pmol) was prepared from bacteriophage T7 as described by Richardson (15 J. Molec. Biol. 49, 1966). ^3H-labeled single-stranded T7 DNA was prepared immediately prior to use by denaturation of doubl - stranded ^3H-labeled T7 DNA with 50 mM NaOH at 20° C for 15 min, followed by neutralization with HCl. The standard exonuclease assay used is a modification of the procedure described by Chase et al. (supra). The standard reaction mixture (100 µl final volume) contained 40 mM Tris/HCl pH 7.5, 10 mM $MgCl_2$, 10 mM dithiothreitol, 60 nmol ^3H-labeled single-stranded T7 DNA (6 cpm/pm), and varying amounts of T7 DNA polymerase. ^3H-labeled double-stranded T7 DNA can also be used as a substrate. Also, any uniformly radioactively labeled DNA, single- or double-stranded, can be used for the assay. Also, 3′ end labeled single-or double-stranded DNA can be used for the assay. After incubation at 37° C for 15 min, the reaction is stopped by the addition of 30 µl of BSA (10mg/ml) and 25 µl of TCA (100% w/v). The assay can be run at 10° C-45° C for 1-60 min. The DNA is precipitated on ice for 15 min (1 min - 12 hr), then centrifuged at 12,000 g for 30 min (5 min - 3 hr). 100 µl of the supernatant is used to det rmine the acid-soluble radioactivity by adding it to 400 µl wat r and 5 ml of aqueous scintillation cocktail.

One unit of exonuclease activity catalyzes the acid solubilization of 10 nmol of total nucleotide in 30 min under the conditions of the assay. Nativ T7 DNA polymerase has a specific exonuclease activity of 5000 units/mg, using the standard assay conditions stated above. The specific exonucl ase activity of the

modified T7 DNA polymerase depends upon the extent of chemical modification, but ideally is at least 10-100-fold lower than that of native T7 DNA polymerase, or 500 to 50 or less units/mg using the standard assay conditions stated above. When double stranded substrate is used the exonuclease activity is about 7-fold higher.

Under the conditions outlined, the exonuclease activity decays exponentially, with a half-life of decay of eight hours. Once per day the reaction vessel is mixed to distribute the soluble oxygen, otherwise the reaction will proceed more rapidly at the surface where the concentration of oxygen is higher. Once per day 2.5 mM DTT (0.3 ml of a fresh 250 mM stock to a 30 ml reaction) is added to replenish the oxidized DTT.

After eight hours, the exonuclease activity of T7 DNA polymerase has been reduced 50%, with negligible loss of polymerase activity. The 50% loss may be the result of the complete inactivation of exonuclease activity of half the polymerase molecules, rather than a general reduction of the rate of exonuclease activity in all the molecules. Thus, after an eight hour reaction all the molecules have normal polymerase activity, half the molecules have normal exonuclease activity, while the other half have <0.1% of their original exonuclease activity.

When 50% of the molecules are modified (an eight hour reaction), the enzyme is suitable, although suboptimal, for DNA sequencing. For more optimum quality of DNA sequencing, the reaction is allowed to proceed to greater than 99% modification (having less than 50 units of exonuclease activity), which requires four days.

After four days, the reaction mixture is dialyzed against 2 changes of 250 ml of 20 mM KPO_4 pH 7.4/0.1 mM dithiothreitol/0.1 mM EDTA/50% glycerol to remove the iron. The modified T7 DNA polymerase (~4 mg/ml) is stored at -20° C.

The reaction mechanism for chemical modification of T7 DNA polymerase depends upon reactive oxygen species generated by the presence of reduced transition metals such as Fe^{2+} and oxygen. A possible reaction mechanism for the generation of hydroxyl radicals is outlined below:

(1) $Fe^{2+} + O_2 \rightarrow Fe^{3+} + O_2^{\cdot}$

(2) $2\,O_2^{\cdot} + 2\,H^{+} \rightarrow H_2O_2 + O_2$

(3) $Fe^{2+} + H_2O_2 \rightarrow FE^{3+} + OH^{\cdot} + OH^{-}$

In equation 1, oxidation of the reduced metal ion yields superoxide radical, $O_2^{\cdot}$. The superoxide radical can undergo a dismutation reaction, producing hydrogen peroxide (equation 2). Finally, hydrogen peroxide can react with reduced metal ions to form hydroxyl radicals, $OH^{\cdot}$ (the Fenton reaction, equation 3). The oxidized metal ion is recycled to the reduced form by reducing agents such as dithiothreitol (DTT).

These reactive oxygen species probably inactivate proteins by irreversibly chemically altering specific amino acid residues. Such damage is observed in SDS-PAGE of fragments of gene 5 produced by CNBr or trypsin. Some fragments disappear, high molecular weight cross linking occurs, and some fragments are broken into two smaller fragments.

As previously mentioned, oxygen, a reducing agent (e.g. DTT, 2-mercaptoethanol) and a transition metal (e.g. iron) are essential elements of the modification reaction. The reaction occurs in air, but is stimulated three-fold by use of 100% oxygen. The reaction will occur slowly in the absence of added transition metals due to the presence of trace quantities of transition metals (1-2μM) in most buffer preparations.

As expected, inhibitors of the modification reaction include anaerobic conditions (e.g., N_2) and metal chelators (e.g. EDTA, citrate, nitrilotriacetate). In addition, the enzymes catalase and superoxide dismutase may inhibit the reaction, consistent with the essential role of reactive oxygen species in the generation of modified T7 DNA polymerase.

As an alternative procedure, it is possible to genetically mutate the T7 gene 5 to specifically inactivate the exonuclease domain of the protein. The T7 gene 5 protein purified from such mutants is ideal for use in DNA sequencing without the need to chemically inactivate the exonuclease by oxidation and without the secondary damage that inevitably occurs to the protein during chemical modification.

Genetically modified T7 DNA polymerase can be isolated by randomly mutagenizing the gene 5 and then screening for those mutants that have lost exonuclease activity, without loss of polymerase activity. Mutagenesis is performed as follows. Single-stranded DNA containing gene 5 (e.g., cloned in pEMBL-8, a plasmid containing an origin for single stranded DNA replication) under the control of a T7 RNA polymerase promoter is prepared by standard procedure, and treated with two different chemical mutagens: hydrazine which will mutat C's and T's, and formic acid, which will mutate G's and A's. Myers et al. 229 Science 242, 1985. The DNA is mutagenized at a dose which results in an average of one base being alter d per plasmid molecule. The single-stranded mutagenized plasmids are then primed with a universal 17-mer primer (see above), and used as templates to synthesize the opposite strands. The synthesiz d strands contain randomly incorporated bases at positions corresponding to the mutated bases in the templates. The double-

stranded mutagenized DNA is then used to transform the strain K38/pGP1-2, which is strain K38 containing the plasmid pGP1-2 (Tabor et al., supra). Upon heat induction this strain expresses T7 RNA polymerase. The transformed cells are plated at 30° C, with approximately 200 colonies per plate.

Screening for cells having T7 DNA polymerase lacking exonuclease activity is based upon the following finding. The 3′ to 5′ exonuclease of DNA polymerases serves a proofreading function. When bases ar misincorporated, the exonuclease will remove the newly incorporated base which is recognized as "abnormal". This is the case for the analog of dATP, etheno-dATP, which is readily incorporated by T7 DNA polymerase in place of dATP. However, in the presence of the 3′ to 5′ exonuclease of T7 DNA polymerase, it is excised as rapidly as it is incorporated, resulting in no net DNA synthesis. As shown in figure 6, using the alternating copolymer poly d(AT) as a template, native T7 DNA polymerase catalyzes extensive DNA synthesis only in the presence of dATP, and not etheno-dATP. In contrast, modified T7 DNA polymerase, because of its lack of an associated exonuclease, stably incorporates etheno-dATP into DNA at a rate comparable to dATP. Thus, using poly d(AT) as a template, and dTTP and etheno-dATP as precursors, native T7 DNA polymerase is unable to synthesize DNA from this template, while T7 DNA polymerase which has lost its exonuclease activity will be able to use this template to synthesize DNA.

The procedure for lysing and screening large number of colonies is described in Raetz (72 Proc. Nat. Acad. Sci. 2274, 1975). Briefly, the K38/pGP1-2 cells transformed with the mutagenized gene 5-containing plasmids are transferred from the petri dish, where they are present at approximately 200 colonies per plate, to a piece of filter paper ("replica plating"). The filter paper discs are then placed at 42° C for 60 min to induce the T7 RNA polymerase, which in turn expresses the gene 5 protein. Thioredoxin is constitutively produced from the chromosomal gene. Lysozyme is added to the filter paper to lyse the cells. After a freeze thaw step to ensure cell lysis, the filter paper discs are incubated with poly d(AT), [α^{32}P]dTTP and. etheno-dATP at 37° C for 60 min. The filter paper discs are then washed with acid to remove the unincorporated [^{32}P]dATP. DNA will precipitate on the filter paper in acid, while nucleotides will be soluble. The washed filter paper is then used to expose X-ray film. Colonies which have induced an active T7 DNA polymerase which is deficient in its exonuclease will have incorporated acid-insoluble ^{32}P, and will be visibl by autoradiography. Colonies expressing native T7 DNA polymerase, or expressing a T7 DNA polymerase defective in polymerase activity, will not appear on the autoradiograph.

Colonies which appear positive are recovered from the master petri dish containing the original colonies. Cells containing each potential positive clone will be induced on a larger scale (one liter) and T7 DNA polymerase purified from each preparation to ascertain the levels of exonuclease associated with each mutant. Those low in exonuclease are appropriate for DNA sequencing.

Directed mutagenesis may also be used to isolate genetic mutants in the exonuclease domain of the T7 gene 5 protein. The following is an example of this procedure.

T7 DNA polymerase with reduced exonuclease activity (modified T7 DNA polymerase) can also be distinguished from native T7 DNA polymerase by its ability to synthesize through regions of secondary structure. Thus, with modified DNA polymerase, DNA synthesis from a labeled primer on a template having secondary structure will result in significantly longer extensions, compared to unmodified or native DNA polymerase. This assay provides a basis for screening for the conversion of small percentages of DNA polymerase molecules to a modified form.

The above assay was used to screen for altered T7 DNA polymerase after treatment with a number of chemical reagents. Three reactions resulted in conversion of the enzyme to a modified form. The first is treatment with iron and a reducing agent, as described above. The other two involve treatment of the enzyme with photooxidizing dyes, Rose Bengal and methylene blue, in the presence of light. The dyes must be titrated carefully, and even under optimum conditions the specificity of inactivation of exonuclease activity over polymerase activity is low, compared to the high specificity of the iron-induced oxidation. Sinc these dyes are quite specific for modification of histidine residues, this result strongly implicates histidine residues as an essential species in the exonuclease active site.

There are 23 histidine residues in T7 gene 5 protein. Eight of these residues lie in the amino half of th protein, in the region where, based on the homology with the large fragment of E. coli DNA polymerase I, the exonuclease domain may be located (Ollis et al. Nature 313, 818. 1984). As described below, seven of th eight histidine residues were mutated individually by synthesis of appropriate oligonucleotides, which were then incorporated into gene 5. These correspond to mutants 1, and 6-10 in table 1.

The mutations wer constructed by first cloning the T7 gene 5 from pGP5-3 (Tabor et al., J. Biol. Chem. 282, 1987) into the SmaI and HindIII sites of the vector M13 mp18, to give mGP5-2. (The vector used and the source of gene 5 are not critical in this procedure.) Single-stranded mGP5-2 DNA was prepared from a strain that incorporates d oxyuracil in place of deoxythymidine (Kunkel, Proc. Natl. Acad. Sci. USA 82, 488, 1985). This procedure provides a strong selection for survival of only the synthesized

strand (that containing the mutation) when transfected into wild-type E.coli, since the strand containing uracil will be preferentially degraded.

Mutant oligonucleotides, 15-20 bases in length, were synthesized by standard procedures. Each oligonucleotide was annealed to the template extended using native T7 DNA polymerase and ligated using T4 DNA ligase. Covalently closed circular molecules were isolated by agarose gel electrophoresis run in the presence of 0.5µg/ml ethidium bromide. The resulting purified molecules were then used to transform E. coli 71.18. DNA from the resulting plaques was isolated and the relevant region sequenced to confirm each mutation.

The following summarizes the oligonucleotides used to generate genetic mutants in the gene 5 exonuclease. The mutations created are underlined. Amino acid and base pair numbers are taken from Dunn et al., 166 J. Molec. Biol. 477, 1983. The relevant wild type sequences of the region of gene 5 mutated are also shown.

Wild type sequence:

```
109 (aa)                                          122 123
Leu Leu Arg Ser Gly Lys Leu Pro Gly Lys Arg Phe Gly Ser His Ala Leu Glu
CTT CTG CGT TCC GGC AAG TTG CCC GGA AAA CGC TTT GGG TCT CAC GCT TTG GAG
14677 (T7 bp)
```

Mutation 1: His 123 → Ser 123

Primer used: 5' CGC TTT GGA TCC TCC GCT TTG 3'

Mutant sequence:

```
                                                    123
Leu Leu Arg Ser Gly Lys Leu Pro Gly Lys Arg Phe Gly Ser Ser Ala Leu Glu
CTT CTG CGT TCC GGC AAG TTG CCC GGA AAA CGC TTT GGA TCC TCC GCT TTG GAG
```

Mutation 2: Deletion of Ser 122 and His 123

Primer used: 5' GGA AAA CGC TTT GGC GCC TTG GAG GCG 3'

Δ

6 base deletion

Mutant sequence:

```
                                                  122 123
Leu Leu Arg Ser Gly Lys Leu Pro Gly Lys Arg Phe Gly ... ... Ala Leu Glu
CTT CTG CGT TCC GGC AAG TTG CCC GGA AAA CGC TTT GGC --- --- GCC TTG GAG
```

Mutati n 3: Ser 122, His 123 → Ala 122, Glu 123

Primer used: 5' CGC TTT GGG GCT GAG GCT TTG G 3'

Mutant sequence:

122 123
Leu Leu Arg Ser Gly Lys Leu Pro Gly Lys Arg Phe Gly ***Ala Glu*** *Ala Leu Glu*
CTT CTG CGT TCC GGC AAG TTG CCC GGA AAA CGC TTT GGG **GCT GAG** GCT TTG GAG

Mutation 4: Lys 118, Arg 119 → Glu 118, Glu 119

Primer used: 5' 5' G CCC GGG GAA GAG TTT GGG TCT CAC GC 3'

Mutant sequence:

118 119
Leu Leu Arg Ser Gly Lys Leu Pro Gly ***Glu Glu*** *Phe Gly Ser His Ala Leu Glu*
CTT CTG CGT TCC GGC AAG TTG CCC GGG **GAA GAG** TTT GGG TCT CAC GCT TTG GAG

Mutation 5:. Arg 111, Ser 112, Lys 114 → Glu 111, Ala 112, Glu 114

Primer used : 5' G GGT CTT CTG **GAA** GCC GGC GAG TTG CCC GG 3'

Mutant sequence:

111 112 114
Leu Leu ***Glu Ala*** *Gly* ***Glu*** *Leu Pro Gly Lys Arg Phe Gly Ser His Ala Leu Glu*
CTT CTG **GAA** GCC GGC GAG TTG CCC GGA AAA CGC TTT GGG TCT CAC GCT TTG GAG

Mutation 6: His 59, His 62 → Ser 59, Ser 62

Primer used: 5' ATT GTG TTC TCC AAC GGA TCC AAG TAT GAC G 3'

Wild-type sequence:

aa: 55 59 62
Leu Ile Val Phe His Asn Gly His Lys Tyr Asp Val
CTT ATT GTG TTC CAC AAC GGT CAC AAG TAT GAC GTT
T7 bp: 14515

Mutant sequence:

59 62
Leu Ile Val Phe ***Ser*** *Asn Gly* ***Ser*** *Lys Tyr Asp Val*
CTT ATT GTG TTC **TCC** AAC GGA **TCC** AAG TAT GAC GTT

Mutation 7: His 82 → Ser 82

Primer used: 5' GAG TTC TCC CTT CCT CG 3'

Wild-type sequence:

aa: 77 82
Leu Asn Arg Glu Phe His Leu Pro Arg Glu Asn
TTG AAC CGA GAG TTC CAC CTT CCT CGT GAG AAC
T7 bp: 14581

Mutant sequence:

82
Leu Asn Arg Glu Phe Ser Leu Pro Arg Glu Asn
TTG AAC CGA GAG TTC TCC CTT CCT CGT GAG AAC

Mutation 8: Arg 96, His 99 → Leu 96, Ser 99

Primer used: 5' CTG TTG ATT TCT TCC AAC CTC 3'

Wild-type sequence:

aa: 93 96 99
Val Leu Ser Arg Leu Ile His Ser Asn Leu Lys Asp Thr Asp
GTG TTG TCA CGT TTG ATT CAT TCC AAC CTC AAG GAC ACC GAT
T7 bp: 14629

Mutant sequence:

96 99
Val Leu Ser Leu Leu Ile Ser Ser Asn Leu Lys Asp Thr Asp
GTG TTG TCA CTG TTG ATT TCT TCC AAC CTC AAG GAC ACC GAT

Mutation 9: His 190 → Ser 190

Primer used: 5' CT GAC AAA TCT TAC TTC CCT 3'

Wild-type sequence:

aa: 185 190
Leu Leu Ser Asp Lys His Tyr Phe Pro Pro Glu
CTA CTC TCT GAC AAA CAT TAC TTC CCT CCT GAG
T7 bp: 14905

Mutant sequence:

190
Leu Leu Ser Asp Lys Ser Tyr Phe Pro Pro Glu
CTA CTC TCT GAC AAA TCT TAC TTC CCT CCT GAG

Mutation 10: His 218 → Ser 218

Primer used: 5' GAC ATT GAA TCT CGT GCT GC 3'

Wild-type sequence:

```
        aa: 214                 218
            Val Asp Ile Glu His Arg Ala Ala Trp Leu Leu
            GTT GAC ATT GAA CAT CGT GCT GCA TGG CTG CTC
    T7 bp: 14992
```

Mutant sequence:

```
                        218
            Val Asp Ile Glu Ser Arg Ala Ala Trp Leu Leu
            GTT GAC ATT GAA TCT CGT GCT GCA TGG CTG CTC
```

Mutation 11: Deletion of amino acids 118 to 123

Primer used: 5' C GGC AAG TTG CCC GGG GCT TTG GAG GCG TGG G 3'

Δ

18 base deletion

Wild-type sequence:

```
109 (aa)                                118              122 123          126
Leu Leu Arg Ser Gly Lys Leu Pro Gly Lys Arg Phe Gly Ser His Ala Leu Glu
CTT CTG CGT TCC GGC AAG TTG CCC GGA AAA CGC TTT GGG TCT CAC GCT TTG GAG
14677 (T7 bp)
```

Mutant sequence:

```
                                    117                          124
Leu Leu Arg Ser Gly Lys Leu Pro Gly·····(6 amino acids)·····Ala Leu Glu
CTT CTG CGT TCC GGC AAG TTG CCC GGG·····(18 bases)·····GCT TTG GAG
```

Mutation 12: His 123 → Glu 123

Primer used: 5' GGG TCT GAG GCT TTG G 3'

Mutant sequence:

```
                                                        123
Leu Leu Arg Ser Gly Lys Leu Pro Gly Lys Arg Phe Gly Ser Glu Ala Leu Glu
CTT CTG CGT TCC GGC AAG TTG CCC GGA AAA CGC TTT GGG TCT GAG GCT TTG GAG
```

Mutation 13: (Arg 131, Lys 136, Lys 140, Lys 144, Arg 145 → Glu 131, Glu 136, Glu 140, Glu 144, Glu 145)

Primer used: 5' GGT TAT GAG CTC GGC GAG ATG GAG GGT GAA TAC GAA GAC GAC TTT GAG GAA ATG CTT GAA G 3'

Wild-type sequence:

```
129(aa) 131                 136                 140                 144 145
Gly Tyr Arg Leu Gly Glu Met Lys Gly Glu Tyr Lys Asp Asp Phe Lys Arg Met Leu Glu Glu
GGT TAT CGC TTA GGC GAG ATG AAG GGT GAA TAC AAA GAC GAC TTT AAG CGT ATG CTT GAA G
14737 (T7 bp)
```

Mutant sequence:

```
129(aa) 131                 136                 140                 144 145
Gly Tyr Glu Leu Gly Glu Met Glu Gly Glu Tyr Glu Asp Asp Phe Glu Glu Met Leu Glu Glu
GGT TAT GAG CTC GGC GAG ATG GAG GGT GAA TAC GAA GAC GAC TTT GAG GAA ATG CTT GAA G
14737 (T7 bp)
```

Each mutant gene 5 protein was produced by infection of the mutant phage into K38/pGP1-2, as follows. The cells were grown at 30° C to an $A_{590} = 1.0$. The temperature was shifted to 42° C for 30 min., to induce T7 RNA polymerase. IPTG was added to 0.5 mM, and a lysate of each phage was added at a moi = 10. Infected cells were grown at 37° C for 90 min. The cells were then harvested and extracts prepared by standard procedures for T7 gene 5 protein.

Extracts were partially purified by passage over a phosphocellulose and DEAE A-50 column, and assayed by measuring the polymerase and exonuclease activities directly, as described above. The results are shown in Table 1.

Table 1
SUMMARY OF EXONUCLEASE AND POLYMERASE ACTIVITIES OF T7 GENE 5 MUTANTS

Mutant	Exonuclease activity, %	Polymerase activity, %
[Wild-type]	[100][a]	[100][b]
Mutant 1 (His 123 → Ser 123)	10-25	>90
Mutant 2 (Δ Ser 122, His 123)	0.2-0.4	>90
Mutant 3 (Ser 122, His 123 → Ala 122, Glu 123)	<2	>90

Table 1
SUMMARY OF EXONUCLEASE AND POLYMERASE ACTIVITIES OF T7 GENE 5 MUTANTS

Mutant	Exonuclease activity, %	Polymerase activity, %
Mutant 4 (Lys 118, Arg 119 → Glu 118, Glu 119)	<30	>90
Mutant 5 (Arg 111, Ser 112, Lys 114 → Glu 111, Ala 112, Glu 114)	>75	>90
Mutant 6 (His 59, His 62 → Ser 59, Ser 62)	>75	>90
Mutant 7 (His 82 → Ser 82)	>75	>90
Mutant 8 (Arg 96, His 99 → Leu 96, Ser 99)	>75	>90
Mutant 9 (His 190 → Ser 190)	>75	>90
Mutant 10 (His 218 → Ser 218)	>75	>90
Mutant 11 (Δ Lys 118, Arg 119, Phe 120, Gly 121, Ser 122, His 123)	<0.02	>90
Mutant 12 (His 123 → Glu 123)	<30	>90
Mutant 13 (Arg 131, Lys 136, Lys 140, Lys 144, Arg 145 → Glu 131, Glu 136, Glu 140, Glu 144, Glu 145)	<30	>90

a. Exonuclease activity was measured on single stranded [^{3}H]T7 DNA. 100% exonuclease activity corresponds to 5,000 units/mg.

b. Polymerase activity was measured using single-stranded calf thymus DNA. 100% polymerase activity corresponds to 8,000 units/mg.

Of the seven histidines tested, only one (His 123: mutant 1) has the enzymatic activities characteristic of modified T7 DNA polymerase. T7 gene 5 protein was purified from this mutant using DEAE-cellulose, phosphocellulose, DEAE-Sephadex and hydroxylapatite chromatography. While the polymerase activity was nearly normal (>90% the level of the native enzyme), the exonuclease activity was reduced 4 to 10-fold.

A variant of this mutant was constructed in which both His 123 and Ser 122 were deleted. The gene 5 protein purified from this mutant has a 200-500 fold lower exonuclease activity, again with retention of >90% of the polymerase activity.

These data strongly suggest that His 123 lies in the active site of the exonuclease domain of T7 gene 5 protein. Furthermore, it is likely that the His 123 is in fact the residue being modified by the oxidation involving iron, oxygen and a reducing agent, since such oxidation has been shown to modify histidine residues in other proteins (Levine, J. Biol. Chem. 258: 11823, 1983; and Hodgson et al. Biochemistry 14:

5294, 1975). The level of residual exonuclease in mutant 11 is comparable to the levels obtainable by chemical modification.

Although mutations at His residues are described, mutations at nearby sites or even at distant sites may also produce mutant enzymes suitable in this invention, e.g., lys and arg (mutants 4 and 15). Similarly, although mutations in some His residues have little effect on exonuclease activity that does not necessarily indicate that mutations near these residues will not affect exonuclease activity. Mutations which are especially effective include those having deletions of 2 or more amino acids, preferably 6-8, for example, near the His-123 region. Other mutations should reduce exonuclease activity further, or completely.

As an example of the use of these mutant strains the following is illustrative. A pGP5-6 (mutation 11)-containing strain has been deposited with the ATCC (see below). The strain is grown as described above and induced as described in Taber et al. J. Biol. Chem. 262:16212 (1987). K38/pTrx-2 cells may be added to increase the yield of genetically modified T7 DNA polymerase.

The above noted deposited strain also contains plasmid pGP1-2 which expresses T7 RNA polymerase. This plasmid is described in Tabor et al., Proc. Nat. Acad. Sci. USA 82:1074, 1985 and was deposited with the ATCC on March 22, 1985 and assigned the number 40,175.

Referring to Fig. 10, pGP5-6 includes the following segments:

1. EcoRI-SacI-SmaI-BamHI polylinker sequence from M13 mp10 (21bp).
2. T7 bp 14309 to 16747, that contains the T7 gene 5, with the following modifications:

T7 bp 14703 is changed from an A to a G, creating a SmaI site.
T7 bp 14304 to 14321 inclusive are deleted (18 bp).

3. SalI-PstI-HindIII polylinker sequence from M13 mp 10 (15 bp)
4. pBR322 bp 29 (HindIII site) to pBR322 bp 375 (BamHI site).
5. T7 bp 22855 to T7 bp 22927, that contains the T7 RNA Polymerase promoter ϕ10, with BamHI linkers inserted at each end (82 bp).
6. pBR322 bp 375 (BamHI site) to pBR322 bp 4361 (EcoRI site).

DNA Sequencing Using Modified T7-type DNA Polymerase

DNA synthesis reactions using modified T7-type DNA polymerase result in chain-terminated fragments of uniform radioactive intensity, throughout the range of several bases to thousands of bases in length. There is virtually no background due to terminations at sites independent of chain terminating agent incorporation (i.e. at pause sites or secondary structure impediments).

Sequencing reactions using modified T7-type DNA polymerase consist of a pulse and chase. By pulse is meant that a short labelled DNA fragment is synthesized; by chase is meant that the short fragment is lengthened until a chain terminating agent is incorporated. The rationale for each step differs from conventional DNA sequencing reactions. In the pulse, the reaction is incubated at 0° C-37° C for 0.5-4 min in the presence of high levels of three nucleotide triphosphates (e.g., dGTP, dCTP and dTTP) and limiting levels of one other labelled, carrier-free, nucleotide triphosphate, e.g., [^{35}S] dATP. Under these conditions the modified polymerase is unable to exhibit its processive character, and a population of radioactive fragments will be synthesized ranging in size from a few bases to several hundred bases. The purpose of the pulse is to radioactively label each primer, incorporating maximal radioactivity while using minimal levels of radioactive nucleotides. In this example, two conditions in the pulse reaction (low temperature, e.g., from 0-20° C, and limiting levels of dATP, e.g., from 0.1μM to 1μM) prevent the modified T7-type DNA polymerase from exhibiting its processive character. Other essential environmental components of the mixture will have similar effects, e.g. limiting more than one nucleotide triphosphate or increasing the ionic strength of the reaction. If the primer is already labelled (e.g., by kinasing) no pulse step is required.

In the chase, the reaction is incubated at 45° C for 1-30 min in the presence of high levels (50-500μM) of all four deoxynucleoside triphosphates and limiting levels (1-50μM) of any one of the four chain terminating agents, e.g., dideoxynucleoside triphosphates, such that DNA synthesis is terminated after an average of 50-600 bases. The purpose of the chase is to extend each radioactively labeled primer under conditions of processive DNA synthesis, terminating each extension exclusively at correct sites in four separate reactions using each of the four dideoxynucleoside triphosphates. Two conditions of the chase (high temperature, e.g., from 30-50° C) and high lev ls (above 50μM) of all four deoxynucleoside triphosphates) allow the modified T7-type DNA polymerase to exhibit its proc ssive character for tens of thousands of bas s; thus the same polymerase molecule will synthesize from the primer-template until a dideoxynucleotide is incorporated. At a chase temperature of 45° C synthesis occurs at >700 nucleotides/sec. Thus, for sequencing reactions the chase is complete in less than a second. ssb increases

processivity, for example, when using dITP, or when using low temperatures or high ionic strength, or low levels of triphosphates throughout the sequencing reaction.

Either [α^{35}S]dATP, [α^{32}P]dATP or fluorescently labell d nucleotides can be used in th DNA sequencing reactions with modified T7-type DNA polymerase. If the fluorescent analog is at the 5′ end of the primer, then no pulse step is required.

Two components determine the average extensions of the synthesis reactions. First is the length of time of the pulse reaction. Since the pulse is done in the absence of chain terminating agents, the longer the pulse reaction time, the longer the primer extensions. At 0°C the polymerase extensions average 10 nucleotides/sec. Second is the ratio of deoxyribonucleoside triphosphates to chain terminating agents in the chase reaction. A modified T7-type DNA polymerase does not discriminate against the incorporation of these analogs, thus the average length of extension in the chase is four times the ratio of the deoxynucleoside triphosphate concentration to the chain terminating agent concentration in the chase reaction. Thus, in order to shorten the average size of the extensions, the pulse time is shortened, e.g., to 30 sec. and/or the ratio of chain terminating agent to deoxynucleoside triphosphate concentration is raised in th chase reaction. This can be done either by raising the concentration of the chain terminating agent or lowering the concentration of deoxynucleoside triphosphate. To increase the average length of the extensions, the pulse time is increased, e.g., to 3-4 min; and/or the concentration of chain terminating agent is lowered (e.g., from 20μM to 2μM) in the chase reaction.

Example 2: DNA sequencing using modified T7 DNA polymerase

The following is an example of a sequencing protocol using dideoxy nucleotides as terminating agents.

9μl of single-stranded M13 DNA (mGP1-2, prepared by standard procedures) at 0.7 mM concentration is mixed with 1 μl of complementary sequencing primer (standard universal 17-mer, 0.5 pmole primer / μl) and 2.5 μl 5X annealing buffer (200 mM Tris-HCl, pH 7.5, 50 mM $MgCl_2$) heated to 65°C for 3 min, and slow cooled to room temperature over 30 min. In the pulse reaction, 12.5 μl of the above annealed mix was mixed with 1 μl dithiothreitol 0.1 M, 2 μl of 3 dNTPs (dGTP, dCTP, dTTP) 3 mM each (P.L Biochemicals in TE), 2.5 μl [α^{35}S]dATP, (1500 Ci/mmol, New England Nuclear) and 1 μl of modified T7 DNA polymeras described in Example 1 (0.4 mg/ml, 2500 units/ml, i.e. 0.4 μg, 2.5 units) and incubated at 0°C, for 2 min, after vortexing and centrifuging in a microfuge for 1 sec. The time of incubation can vary from 30 sec to 20 min and temperature can vary from 0°C to 37°C. Longer times are used for determining sequences distant from the primer.

4.5 μl aliquots of the above pulse reaction are added to each of four tubes containing the chase mixes, preheated to 45°C. The four tubes, labeled G, A, T, C, each contain trace amounts of either dideoxy (dd) G, A, T, or C (P-L Biochemicals). The specific chase solutions are given below. Each tube contains 1.5 μl dATP 1mM, 0.5 μl 5X annealing buffer (200 mM Tris-HCl, pH 7.5, 50mM $MgCl_2$), and 1.0 μl ddNTP 100 μM (where ddNTP corresponds to ddG,A,T or C in the respective tubes). Each chase reaction is incubated at 45°C (or 30°C-50°C) for 10 min, and then 6 μl of stop solution (90% formamide, 10mM EDTA, 0.1% xylenecyanol) is added to each tube, and the tube placed on ice. The chase times can vary from 1-30 min.

The sequencing reactions are run on standard, 6% polyacrylamide sequencing gel in 7M urea, at 30 Watts for 6 hours. Prior to running on a gel the reactions are heated to 75°C for 2 min. The gel is fixed in 10% acetic acid, 10% methanol, dried on a gel dryer, and exposed to Kodak OM1 high-contrast autoradiography film overnight.

Example 3: DNA sequencing using limiting concentrations of dNTPS

In this example DNA sequence analysis of mGP1-2 DNA is performed using limiting levels of all four deoxyribonucleoside triphosphates in the pulse reaction. This method has a number of advantages over the protocol in example 2. First, the pulse reaction runs to completion, whereas in the previous protocol it was n cessary to interrupt a time course. As a consequence the reactions ar easier to run. Second, with this method it is easier to control the extent of the elongations in th puls , and so the efficiency of labeling of s quences near the primer (th first 50 bases) is increased approximately 10-fold.

7 μl of 0.75 mM single-stranded M13 DNA (mGP1-2) was mixed with 1μl of compl mentary sequencing primer (17-mer, 0.5 pmole primer/μl) and 2 μl 5X annealing buffer (200 mM Tris-HCl pH 7.5, 50 mM $MgCl_2$, 250 mM NaCl) heated at 65°C for 2 min, and slowly cooled to room temperature over 30 min. In the puls reaction 10 μl of the abov annealed mix was mixed with 1 μl dithiothreitol 0.1 M, 2 μl of 3 dNTPs (dGTP,

dCTP, dTTP) 1.5 µM each, 0.5µl [[α^{35}S]dATP, (α10µM) (about 10µM, 1500 Ci/mmol, New England Nuclear) and 2 µl modified T7 DNA polymerase (0.1 mg/ml, 1000 units/ml, i.e., 0.2 µg, 2 units) and incubated at 37° C for 5 min. (The temperature and time of incubation can be varied from 20° C-45° C and 1-60 min., respectively.)

3.5 µl aliquots of the above pulse reaction were added to each of four tubes containing the chase mixes, which were preheated to 37° C. The four tubes, labeled G, A, T, C, each contain trace amounts of either dideoxy G, A, T, C. The specific chase solutions are given below. Each tube contains 0.5 µl 5X annealing buffer (200 mM Tris-HCl pH 7.5, 50 mM $MgCl_2$, 250 mM NaC_1), 1 µl 4dNTPs (dGTP, dATP, dTTP, dCTP) 200 µM each, and 1.0 µl ddNTP 20 µM. Each chase reaction is incubated at 37° C for 5 min (or 20° C-45° C and 1-60 min respectively), and then 4 µl of a stop solution (95% formamide, 20 mM EDTA, 0.05% xylene-cyanol) added to each tube, and the tube placed on ice prior to running on a standard polyacrylamide sequencing gel as described above.

Example 4: Replacement of dGTP with dITP for DNA sequencing

In order to sequence through regions of compression in DNA, i.e., regions having compact secondary structure, it is common to use dITP (Mills et al., 76 Proc. Natl. Acad. Sci. 2232, 1979) or deazaguanosine triphosphate (deaza GTP, Mizusawa et al., 14 Nuc. Acid Res. 1319, 1986). We have found that both analogs function well with T7-type polymerases, especially with dITP in the presence of ssb. Preferably these reactions are performed with the above described genetically modified T7 polymerase, or the chase reaction is for 1-2 min., and/or at 20° C to reduce exonuclease degradation.

Modified T7 DNA polymerase efficiently utilizes dITP or deoza-GTP in place of dGTP. dITP is substituted for dGTP in both the pulse and chase mixes at a concentration two to five times that at which dGTP is used. In the ddG chase mix ddGTP is still used (not ddITP).

The chase reactions using dITP are sensitive to the residual low levels (about 0.01 units) of exonuclease activity. To avoid this problem, the chase reaction times should not exceed 5 min when dITP is used. It is recommended that the four dITP reactions be run in conjunction with, rather than to the exclusion of, the four reactions using dGTP. If both dGTP and dITP are routinely used, the number of required mixes can be minimized by: (1) Leaving dGTP and dITP out of the chase mixes, which means that the four chase mixes can be used for both dGTP and dITP chase reactions. (2) Adding a high concentration of dGTP or dITP (2µl at 0.5 mM and 1-2.5 mM respectively) to the appropriate pulse mix. The two pulse mixes then each contain a low concentration of dCTP,dTTP and [α^{35}S]dATP, and a high concentration of either dGTP or dITP. This modification does not usually adversely effect the quality of the sequencing reactions, and reduces the required number of pulse and chase mixes to run reactions using both dGTP and dITP to six.

The sequencing reaction is as for example 3, except that two of the pulse mixes contain a) 3 dNTP mix for dGTP: 1.5 µM dCTP,dTTP, and 1 mM dGTP and b) 3 dNTP mix for dITP: 1.5 µM dCTP,dTTP, and 2 mM dITP. In the chase reaction dGTP is removed from the chase mixes (i.e. the chase mixes contain 30 µM dATP,dTTP and dCTP, and one of the four dideoxynucleotides at 8 µM), and the chase time using dITP does not exceed 5 min.

Deposits

Strains K38/pGP5-5/pTrx-2, K38/pTrx-2 and M13 mGP1-2 have been deposited with the ATCC and assigned numbers 67,287, 67,286, and 40,303 respectively. These deposits were made on January 13, 1987. Strain K38/pGP1-2/pGP5-6 was deposited with the ATCC. On December 4, 1987, and assigned the number 67571.

Applicants' and their assignees acknowledge their responsibility to replace these cultures should they die before the end of the term of a patent issued hereon, 5 years after the last request for a culture, or 30 years, whichever is the longer, and its responsibility to notify the depository of the issuance of such a patent, at which time the deposits will be made irrevocably available to the public. Until that time the deposits will be mad irrevocably available to the Commissioner of Patents under the terms of 37 CFR Section 1-14 and 35 USC Section 112.

Other Embodiments

Other embodiments are within the following claims.

Other uses of the modified DNA polymerases of this invention, which take advantag of their processivity, and lack of xonucl ase activity, includ the direct enzymatic amplification of genomic DNA sequences. This has been described, for other polymerases, by Saiki et al., 230 Science 1350, 1985; and Scharf, 233 Science 1076, 1986.

Referring to Fig. 6, enzymatic amplification of a specific DNA region entails the use of two primers which anneal to opposite strands of a double stranded DNA sequence in the region of interest, with their 3′ ends directed toward one another (see dark arrows). The actual procedure involves multiple (10-40, preferably 16-20) cycles of denaturation, annealing, and DNA synthesis. Using this procedure it is possible to amplify a specific region of human genomic DNA over 200,000 times. As a result the specific gene fragment represents about one part in five, rather than the initial one part in a million. This greatly facilitates both the cloning and the direct analysis of genomic DNA. For diagnostic uses, it can speed up the analysis from several weeks to 1-2 days.

Unlike Klenow fragment, where the amplification process is limited to fragments under two hundred bases in length, modified T7-type DNA polymerases should (preferably in conjuction with E. coli DNA binding protein, or ssb, to prevent "snapback formation of single stranded DNA) permit the amplification of DNA fragments thousands of bases in length.

The modified T7-type DNA polymerases are also suitable in standard reaction mixtures: for a) filling in 5′ protruding termini of DNA fragments generated by restriction enzyme cleavage; in order to, for example, produce blunt-ended double stranded DNA from a linear DNA molecule having a single stranded region with no 3′ protruding termini; b) for labeling the 3′ termini of restriction fragments, for mapping mRNA start sites by S1 nuclease analysis, or sequencing DNA using the Maxam and Gilbert chemical modification procedure; and c) for in vitro mutagenesis of cloned DNA fragments. For example, a chemically synthesized primer which contains specific mismatched bases is hybridized to a DNA template, and then extended by the modified T7-type DNA polymerase. In this way the mutation becomes permanently incorporated into the synthesized strand. It is advantageous for the polymerase to synthesize from the primer through the entire length of the DNA. This is most efficiently done using a processive DNA polymerase. Alternatively mutagenesis is performed by misincorporation during DNA synthesis (see above). This application is used to mutagenize specific regions of cloned DNA fragments. It is important that the enzyme used lack exonuclease activity. By standard reaction mixture is meant a buffered solution containing the polymerase and any necessary deoxynucleosides, or other compounds.

Claims

1. A purified modified gene characterized in that it encodes a processive modified DNA polymeras , which has sufficient DNA polymerase activity for use in DNA sequencing when said polymerase is combined with any host factor necessary for said DNA polymerase activity, and which results from the modification of a naturally occurring gene modified to reduce the activity of naturally occurring exonuclease activity of the naturally occurring DNA polymerase.

2. A purified modified gene as claimed in claim 1 further characterized in that said polymerase is a modified bacteriophage T7-type DNA polymerase which has an exonuclease activity at least 50% lower than the naturally-occurring exonuclease activity of naturally occurring T7-type DNA polymerase.

3. A purified modified gene as claimed in claim 2 further characterized in that said gene has been modified to eliminate the naturally occurring exonuclease activity of the naturally occurring DNA polymerase.

4. A purified modified gene as claimed in claim 2 further characterized in that said polymerase lacks one or more amino acids present in native DNA polymerase, and thereby has a reduced exonuclease activity.

5. A purified modified gene as claimed in claim 1 further characterized in that a said amino acid has been replaced by an amino acid other than that naturally occurring at the site of substitution, and the DNA polymerase encoded ther by has a reduced exonuclease activity.

6. A purified modified gene as claimed in claim 2 further charact rized in the said polymerase is T7 DNA polymerase.

7. A purified modified DNA polymerase encoded by the gene of claim 1, 2, 3, 4, 5, or 6.

FIGURE 1

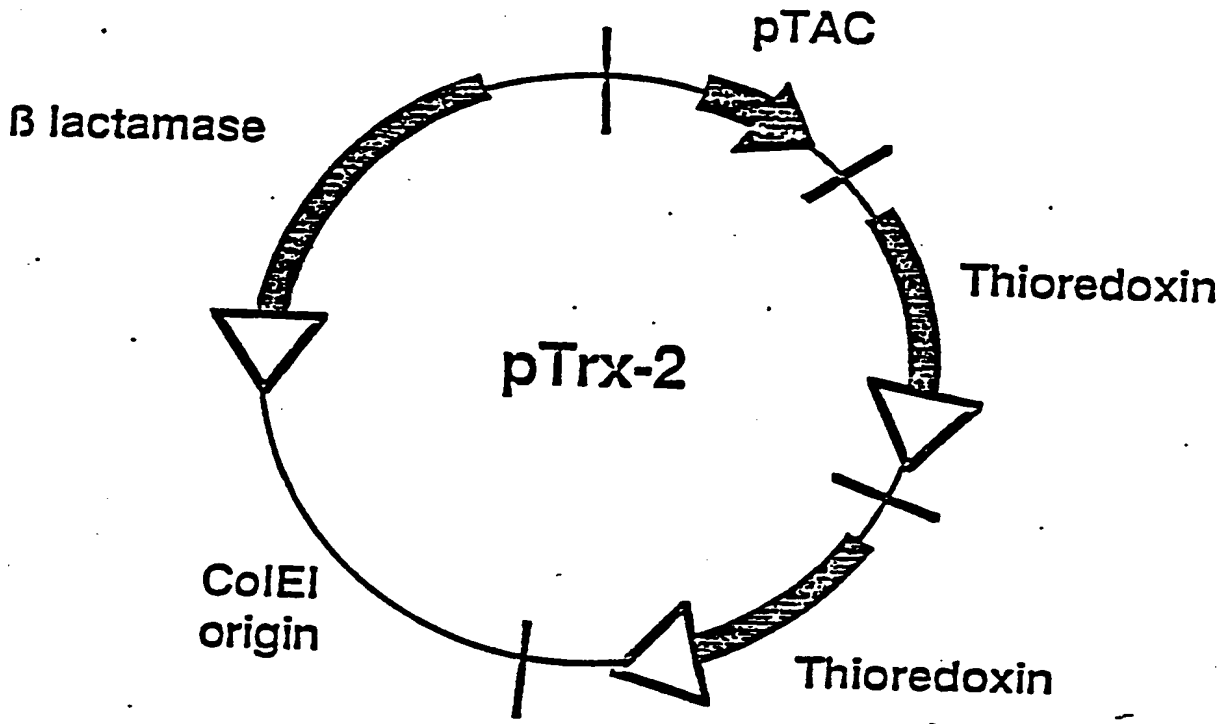

pTAC
ß lactamase
Thioredoxin
pTrx-2
ColEI
origin
Thioredoxin

FIGURE 2

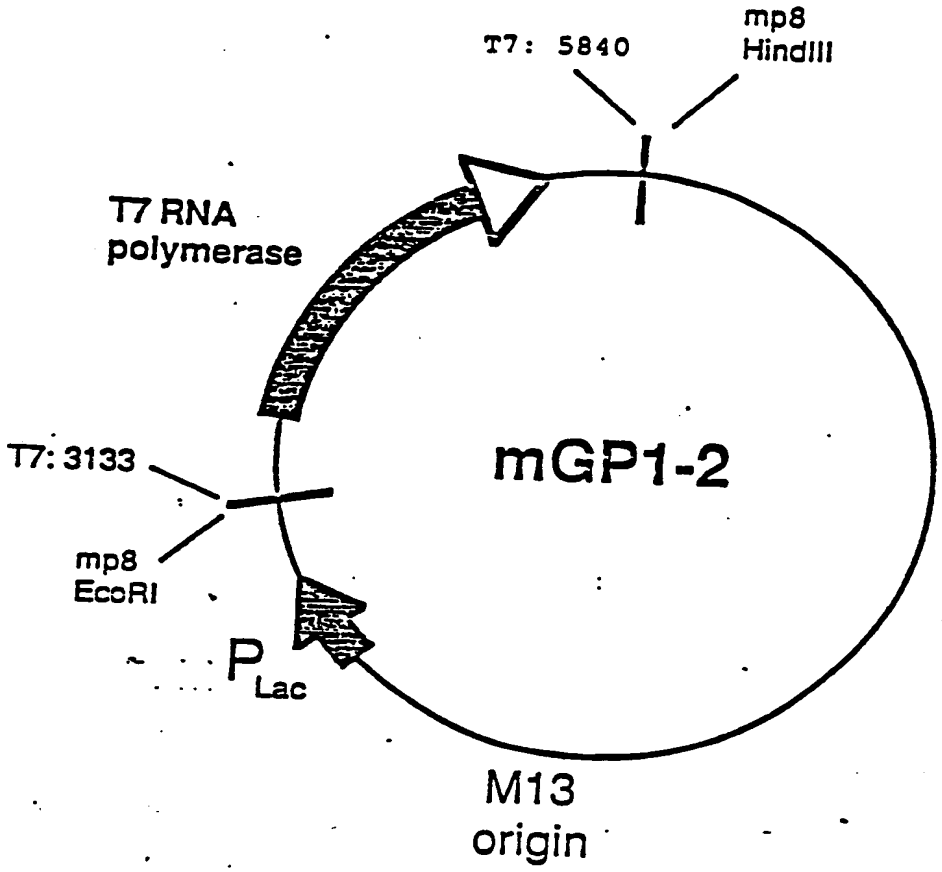

T7: 5840
mp8
HindIII
T7 RNA
polymerase
T7: 3133
mGP1-2
mp8
EcoRI
P Lac
M13
origin

FIGURE 3

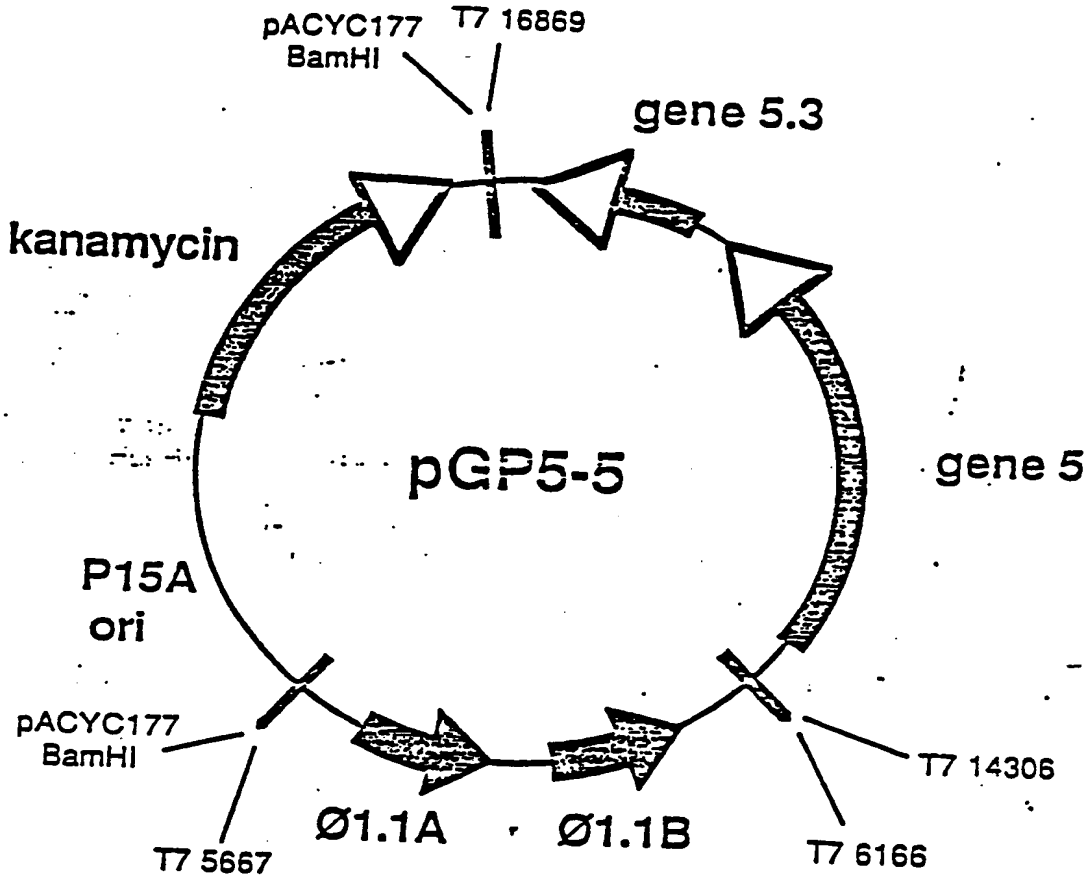

pACYC177
BamHI
T7 16869
gene 5.3
kanamycin
pGP5-5
gene 5
P15A
ori
pACYC177
BamHI
T7 14306
Ø1.1A
Ø1.1B
T7 5667
T7 6166

FIGURE 4

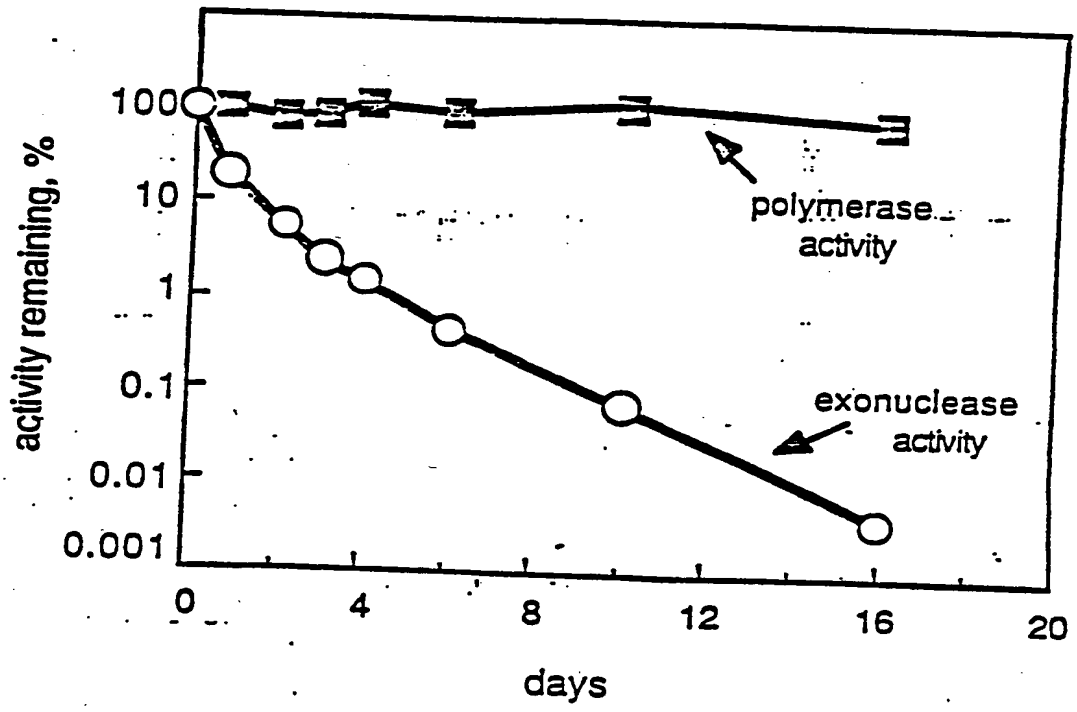
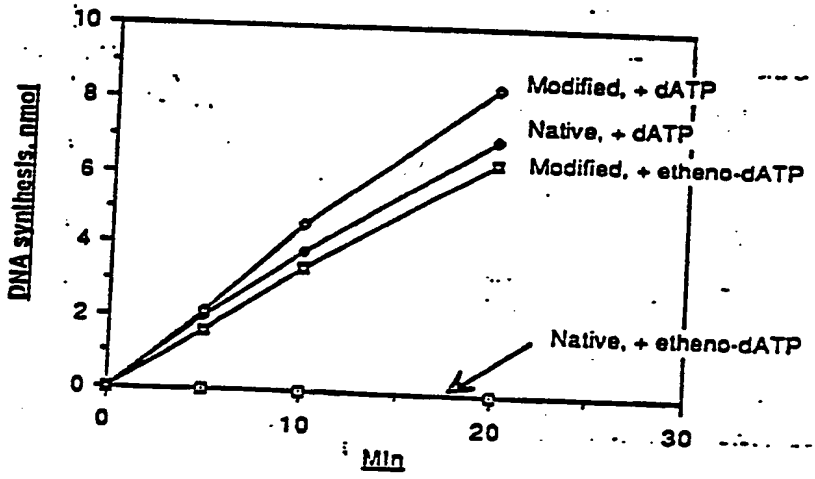

activity remaining, %
100
10
1
0.1
0.01
0.001
0
4
8
12
16
20
days
polymerase activity
exonuclease activity

FIGURE 5


DNA synthesis, nmol
10
8
6
4
2
0
0
10
20
30
Min
Modified, + dATP
Native, + dATP
Modified, + etheno-dATP
Native, + etheno-dATP

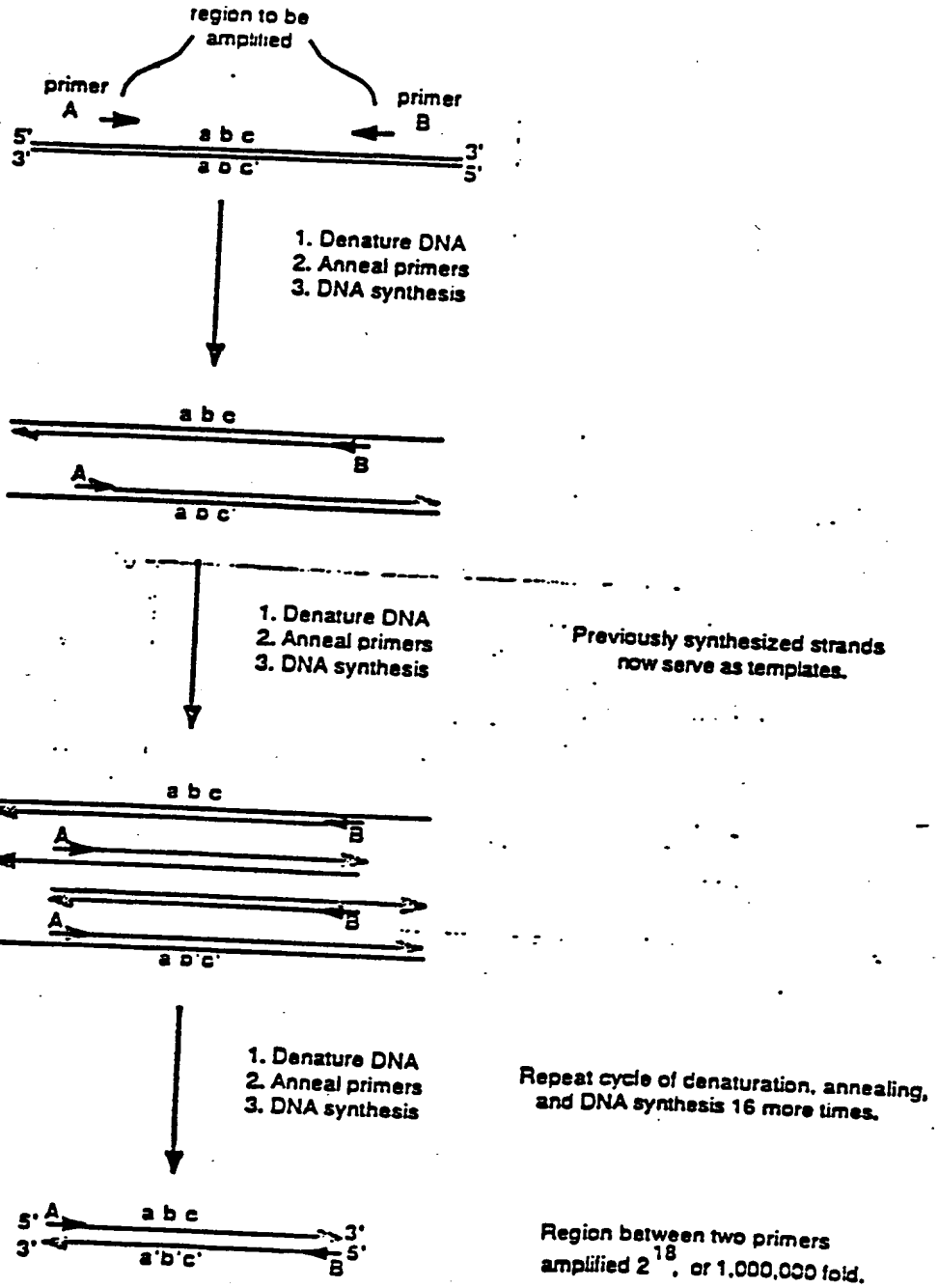

region to be amplified
primer A
primer B
5'
3'
a b c
a b c'
3'
5'
1. Denature DNA
2. Anneal primers
3. DNA synthesis
a b c
A
B
a b c'
1. Denature DNA
2. Anneal primers
3. DNA synthesis
Previously synthesized strands now serve as templates.
a b c
A
B
A
B
a b c'
1. Denature DNA
2. Anneal primers
3. DNA synthesis
Repeat cycle of denaturation, annealing, and DNA synthesis 16 more times.
5'
A
a b c
3'
3'
a'b'c'
5'
B
Region between two primers amplified 2 18 , or 1,000,000 fold.

FIGURE 7

```
        10          20          30          40          50
TTCTTCTCAT GTTTGACAGC TTATCATCGA CTGCACGGTG CACCAATGCT
        60          70          80          90         100
TCTGGCGTCA GGCAGCCATC GGAAGCTGTG GTATGGCTGT GCAGGTCGTA
       110         120         130         140         150
AATCACTGCA TAATTCGTGT CGCTCAAGGC GCACTCCCGT TCTGGATAAT
       160         170         180         190         200
GTTTTTTGCG CCGACATCAT AACGGTTCTG GCAAATATTC TGAAATGAGC
       210         220         230         240         250
TGTTGACAAT TAATCATCGG CTCGTATAAT GTGTGGAATT GTGAGCGGAT
       260         270         280         290         300
AACAATTTCA CACAGGAAAC AGGGGATCCG TCAACCTTTA GTTGGTTAAT
       310         320         330         340         350
GTTACACCAA CAACGAAACC AACACGCCAG GCTTATTCCT GTGGAGTTAT
       360         370         380         390         400
ATATGAGCGA TAAAATTATT CACCTGACTG ACGACAGTTT TGACACGGAT
       410         420         430         440         450
GTACTCAAAG CGGACGGGGC GATCCTCGTC GATTTCTGGG CAGAGTGGTG
       460         470         480         490         500
CGGTCCGTGC AAGATGATCG CCCCGATTCT GGATGAAATC GCTGACGAAT
```

FIGURE 7 (continued)

```
       510         520         530         540         550
ATCAGGGCAA ACTGACCGTT GCAAAACTGA ACATCGATCA AAACCCTGGT
       560         570         580         590         600
ACTGCGCCGA AATATGGCAT CCGTGGTATC CCGACTCTGC TGCTGTTCAA
       610         620         630         640         650
AAACGGTGAA GTGGCGGCAA CCAAAGTGGG TGCACTGTCT AAAGGTCAGT
       660         670         680         690         700
TGAAAGAGTT CCTCGACGCT AACCTGGCGT AAGGGAATTT CATGTTCGGG
       710         720         730         740         750
TGCCCCGTCG CTAAAAACTG GACGCCCGGC GTGAGTCATG CTAACTTAGT
       760         770         780         790         800
GTTGACGGAT CCCCGGGGAT CCGTCAACCT TTAGTTGGTT AATGTTACAC
       810         820         830         840         850
CAACAACGAA ACCAACACGC CAGGCTTATT CCTGTGGAGT TATATATGAG
       860         870         880         890         900
CGATAAAATT ATTCACCTGA CTGACGACAG TTTTGACACG GATGTACTCA
       910         920         930         940         950
AAGCGGACGG GGCGATCCTC GTCGATTTCT GGGCAGAGTG GTGCGGTCCG
       960         970         980         990        1000
TGCAAGATGA TCGCCCCGAT TCTGGATGAA ATCGCTGACG AATATCAGGG
      1010        1020        1030        1040        1050
CAAACTGACC GTTGCAAAAC TGAACATCGA TCAAAACCCT GGTACTGCGC
      1060        1070        1080        1090        1100
CGAAATATGG CATCCGTGGT ATCCCGACTC TGCTGCTGTT CAAAAACGGT
      1110        1120        1130        1140        1150
GAAGTGGCGG CAACCAAAGT GGGTGCACTG TCTAAAGGTC AGTTGAAAGA
      1160        1170        1180        1190        1200
GTTCCTCGAC GCTAACCTGG CGTAAGGGAA TTTCATGTTC GGGTGCCCCG
      1210        1220        1230        1240        1250
TCGCTAAAAA CTGGACGCCC GGCGTGAGTC ATGCTAACTT AGTGTTGACG
      1260        1270        1280        1290        1300
GATCCCCCTG CCTCGCGCGT TTCGGTGATG ACGGTGAAAA CCTCTGACAC
      1310        1320        1330        1340        1350
ATGCAGCTCC CGGAGACGGT CACAGCTTGT CTGTAAGCGG ATGCCGGGAG
      1360        1370        1380        1390        1400
CAGACAAGCC CGTCAGGGCG CGTCAGCGGG TGTTGGCGGG TGTCGGGGCG
      1410        1420        1430        1440        1450
CAGCCATGAC CCAGTCACGT AGCGATAGCG GAGTGTATAC TGGCTTAACT
      1460        1470        1480        1490        1500
ATGCGGCATC AGAGCAGATT GTACTGAGAG TGCACCATAT GCGGTGTGAA
      1510        1520        1530        1540        1550
ATACCGCACA GATGCGTAAG GAGAAAATAC CGCATCAGGC GCTCTTCCGC
      1560        1570        1580        1590        1600
TTCCTCGCTC ACTGACTCGC TGCGCTCGGT CGTTCGGCTG CGGCGAGCGG
      1610        1620        1630        1640        1650
TATCAGCTCA CTCAAAGGCG GTAATACGGT TATCCACAGA ATCAGGGGAT
      1660        1670        1680        1690        1700
AACGCAGGAA AGAACATGTG AGCAAAAGGC CAGCAAAAGG CCAGGAACCG
      1710        1720        1730        1740        1750
TAAAAAGGCC GCGTTGCTGG CGTTTTTCCA TAGGCTCCGC CCCCCTGACG
      1760        1770        1780        1790        1800
AGCATCACAA AAATCGACGC TCAAGTCAGA GGTGGCGAAA CCCGACAGGA
      1810        1820        1830        1840        1850
CTATAAAGAT ACCAGGCGTT TCCCCCTGGA AGCTCCCTCG TGCGCTCTCC
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FIGURE 7 (continued)

```
      1860        1870        1880        1890        1900
TGTTCCGACC CTGCCGCTTA CCGGATACCT GTCCGCCTTT CTCCCTTCGG
      1910        1920        1930        1940        1950
GAAGCGTGGC GCTTTCTCAA TGCTCACGCT GTAGGTATCT CAGTTCGGTG
      1960        1970        1980        1990        2000
TAGGTCGTTC GCTCCAAGCT GGGCTGTGTG CACGAACCCC CCGTTCAGCC
      2010        2020        2030        2040        2050
CGACCGCTGC GCCTTATCCG GTAACTATCG TCTTGAGTCC AACCCGGTAA
      2060        2070        2080        2090        2100
GACACGACTT ATCGCCACTG GCAGCAGCCA CTGGTAACAG GATTAGCAGA
      2110        2120        2130        2140        2150
GCGAGGTATG TAGGCGGTGC TACAGAGTTC TTGAAGTGGT GGCCTAACTA
      2160        2170        2180        2190        2200
CGGCTACACT AGAAGGACAG TATTTGGTAT CTGCGCTCTG CTGAAGCCAG
      2210        2220        2230        2240        2250
TTACCTTCGG AAAAAGAGTT GGTAGCTCTT GATCCGGCAA ACAAACCACC
      2260        2270        2280        2290        2300
GCTGGTAGCG GTGGTTTTTT TGTTTGCAAG CAGCAGATTA CGCGCAGAAA
      2310        2320        2330        2340        2350
AAAAGGATCT CAAGAAGATC CTTTGATCTT TTCTACGGGG TCTGACGCTC
      2360        2370        2380        2390        2400
AGTGGAACGA AAACTCACGT TAAGGGATTT TGGTCATGAG ATTATCAAAA
      2410        2420        2430        2440        2450
AGGATCTTCA CCTAGATCCT TTTAAATTAA AAATGAAGTT TTAAATCAAT
      2460        2470        2480        2490        2500
CTAAAGTATA TATGAGTAAA CTTGGTCTGA CAGTTACCAA TGCTTAATCA
      2510        2520        2530        2540        2550
GTGAGGCACC TATCTCAGCG ATCTGTCTAT TTCGTTCATC CATAGTTGCC
      2560        2570        2580        2590        2600
TGACTCCCCG TCGTGTAGAT AACTACGATA CGGGAGGGCT TACCATCTGG
      2610        2620        2630        2640        2650
CCCCAGTGCT GCAATGATAC CGCGAGACCC ACGCTCACCG GCTCCAGATT
      2660        2670        2680        2690        2700
TATCAGCAAT AAACCAGCCA GCCGGAAGGG CCGAGCGCAG AAGTGGTCCT
      2710        2720        2730        2740        2750
GCAACTTTAT CCGCCTCCAT CCAGTCTATT AATTGTTGCC GGGAAGCTAG
      2760        2770        2780        2790        2800
AGTAAGTAGT TCGCCAGTTA ATAGTTTGCG CAACGTTGTT GCCATTGCTG
      2810        2820        2830        2840        2850
CAGGCATCGT GGTGTCACGC TCGTCGTTTG GTATGGCTTC ATTCAGCTCC
      2860        2870        2880        2890        2900
GGTTCCCAAC GATCAAGGCG AGTTACATGA TCCCCCATGT TGTGCAAAAA
      2910        2920        2930        2940        2950
AGCGGTTAGC TCCTTCGGTC CTCCGATCGT TGTCAGAAGT AAGTTGGCCG
      2960        2970        2980        2990        3000
CAGTGTTATC ACTCATGGTT ATGGCAGCAC TGCATAATTC TCTTACTGTC
      3010        3020        3030        3040        3050
ATGCCATCCG TAAGATGCTT TTCTGTGACT GGTGAGTACT CAACCAAGTC
      3060        3070        3080        3090        3100
ATTCTGAGAA TAGTGTATGC GGCGACCGAG TTGCTCTTGC CCGGCGTCAA
      3110        3120        3130        3140        3150
CACGGGATAA TACCGCGCCA CATAGCAGAA CTTTAAAAGT GCTCATCATT
      3160        3170        3180        3190        3200
GGAAAACGTT CTTCGGGGCG AAAACTCTCA AGGATCTTAC CGCTGTTGAG
```

FIGURE 7 (continued)

```
      3210        3220        3230        3240        3250
ATCCAGTTCG ATGTAACCCA CTCGTGCACC CAACTGATCT TCAGCATCTT
      3260        3270        3280        3290        3300
TTACTTTCAC CAGCGTTTCT GGGTGAGCAA AAACAGGAAG GCAAAATGCC
      3310        3320        3330        3340        3350
GCAAAAAAGG GAATAAGGGC GACACGGAAA TGTTGAATAC TCATACTCTT
      3360        3370        3380        3390        3400
CCTTTTTCAA TATTATTGAA GCATTTATCA GGGTTATTGT CTCATGAGCG
      3410        3420        3430        3440        3450
GATACATATT TGAATGTATT TAGAAAAATA AACAAATAGG GGTTCCGCGC
      3460        3470        3480        3490        3500
ACATTTCCCC GAAAAGTGCC ACCTGACGTC TAAGAAACCA TTATTATCAT
      3510        3520        3530        3540        3550
GACATTAACC TATAAAAATA GGCGTATCAC GAGGCCCTTT CGTCTTCAAG

AA
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FIGURE 8

```
        10          20          30          40          50
GTTGACACAT ATGAGTCTTG TGATGTACTG GCTGATTTCT ACGACCAGTT
        60          70          80          90          100
CGCTGACCAG TTGCACGAGT CTCAATTGGA CAAAATGCCA GCACTTCCGG
        110         120         130         140         150
CTAAAGGTAA CTTGAACCTC CGTGACATCT TAGAGTCGGA CTTCGCGTTC
        160         170         180         190         200
GCGTAACGCC AAATCAATAC GACTCACTAT AGAGGGACAA ACTCAAGGTC
        210         220         230         240         250
ATTCGCAAGA GTGGCCTTTA TGATTGACCT TCTTCCGGTT AATACGACTC
        260         270         280         290         300
ACTATAGGAG AACCTTAAGG TTTAACTTTA AGACCCTTAA GTGTTAATTA
        310         320         330         340         350
GAGATTTAAA TTAAAGAATT ACTAAGAGAG GACTTTAAGT ATGCGTAACT
        360         370         380         390         400
TCGAAAAGAT GACCAAACGT TCTAACCGTA ATGCTCGTGA CTTCGAGGCA
        410         420         430         440         450
ACCAAAGGTC GCAAGTTGAA TAAGACTAAG CGTGACCGCT CTCACAAGCG
        460         470         480         490         500
TAGCTGGGAG GGTCAGTAAG ATGGGACGTT TATATAGTGG TAATCTGGCA
        510         520         530         540         550
CCGGATCCGG TATGAAGAGA TTGTTAAGTC ACGATAATCA ATAGGAGAAA
        560         570         580         590         600
TCAATATGAT CGTTTCTGAC ATCGAAGCTA ACGCCCTCTT AGAGAGCGTC
```

FIGURE 8 (continued)

```
       610         620         630         640         650
ACTAAGTTCC ACTGCGGGGT TATCTACGAC TACTCCACCG CTGAGTACGT
       660         670         680         690         700
AAGCTACCGT CCGAGTGACT TCGGTGCGTA TCTGGATGCG CTGGAAGCCG
       710         720         730         740         750
AGGTTGCACG AGGCGGTCTT ATTGTGTTCC ACAACGGTCA CAAGTATGAC
       760         770         780         790         800
GTTCCTGCAT TGACCAAACT GGCAAAGTTG CAATTGAACC GAGAGTTCCA
       810         820         830         840         850
CCTTCCTCGT GAGAACTGTA TTGACACCCT TGTGTTGTCA CGTTTGATTC
       860         870         880         890         900
ATTCCAACCT CAAGGACACC GATATGGGTC TTCTGCGTTC CGGCAAGTTG
       910         920         930         940         950
CCCGGAAAAC GCTTTGGGTC TCACGCTTTG GAGGCGTGGG GTTATCGCTT
       960         970         980         990        1000
AGGCGAGATG AAGGGTGAAT ACAAAGACGA CTTTAAGCGT ATGCTTGAAG
      1010        1020        1030        1040        1050
AGCAGGGTGA AGAATACGTT GACGGAATGG AGTGGTGGAA CTTCAACGAA
      1060        1070        1080        1090        1100
GAGATGATGG ACTATAACGT TCAGGACGTT GTGGTAACTA AAGCTCTCCT
      1110        1120        1130        1140        1150
TGAGAAGCTA CTCTCTGACA AACATTACTT CCCTCCTGAG ATTGACTTTA
      1160        1170        1180        1190        1200
CGGACGTAGG ATACACTACG TTCTGGTCAG AATCCCTTGA GGCCGTTGAC
      1210        1220        1230        1240        1250
ATTGAACATC GTGCTGCATG GCTGCTCGCT AAACAAGAGC GCAACGGGTT
      1260        1270        1280        1290        1300
CCCGTTTGAC ACAAAAGCAA TCGAAGAGTT GTACGTAGAG TTAGCTGCTC
      1310        1320        1330        1340        1350
GCCGCTCTGA GTTGCTCCGT AAATTGACCG AAACGTTCGG CTCGTGGTAT
      1360        1370        1380        1390        1400
CAGCCTAAAG GTGGCACTGA GATGTTCTGC CATCCGCGAA CAGGTAAGCC
      1410        1420        1430        1440        1450
ACTACCTAAA TACCCTCGCA TTAAGACACC TAAAGTTGGT GGTATCTTTA
      1460        1470        1480        1490        1500
AGAAGCCTAA GAACAAGGCA CAGCGAGAAG GCCGTGAGCC TTGCGAACTT
      1510        1520        1530        1540        1550
GATACCCGCG AGTACGTTGC TGGTGCTCCT TACACCCCAG TTGAACATGT
      1560        1570        1580        1590        1600
TGTGTTTAAC CCTTCGTCTC GTGACCACAT TCAGAAGAAA CTCCAAGAGG
      1610        1620        1630        1640        1650
CTGGGTGGGT CCCGACCAAG TACACCGATA AGGGTGCTCC TGTGGTGGAC
      1660        1670        1680        1690        1700
GATGAGGTAC TCGAAGGAGT ACGTGTAGAT GACCCTGAGA AGCAAGCCGC
      1710        1720        1730        1740        1750
TATCGACCTC ATTAAAGAGT ACTTGATGAT TCAGAAGCGA ATCGGACAGT
      1760        1770        1780        1790        1800
CTGCTGAGGG AGACAAAGCA TGGCTTCGTT ATGTTGCTGA GGATGGTAAG
      1810        1820        1830        1840        1850
ATTCATGGTT CTGTTAACCC TAATGGAGCA GTTACGGGTC GTGCGACCCA
      1860        1870        1880        1890        1900
TGCGTTCCCA AACCTTGCGC AAATTCCGGG TGTACGTTCT CCTTATGGAG
      1910        1920        1930        1940        1950
AGCAGTGTCG CGCTGCTTTT GGCGCTGAGC ACCATTTGGA TGGGATAACT
```

FIGURE 8 (continued)

```
      1960        1970        1980        1990        2000
GGTAAGCCTT GGGTTCAGGC TGGCATCGAC GCATCCGGTC TTGAGCTACG
      2010        2020        2030        2040        2050
CTGCTTGGCT CACTTCATGG CTCGCTTTGA TAACGGCGAG TACGCTCACG
      2060        2070        2080        2090        2100
AGATTCTTAA CGGCGACATC CACACTAAGA ACCAGATAGC TGCTGAACTA
      2110        2120        2130        2140        2150
CCTACCCGAG ATAACGCTAA GACGTTCATC TATGGGTTCC TCTATGGTGC
      2160        2170        2180        2190        2200
TGGTGATGAG AAGATTGGAC AGATTGTTGG TGCTGGTAAA GAGCGCGGTA
      2210        2220        2230        2240        2250
AGGAACTCAA GAAGAAATTC CTTGAGAACA CCCCCGCGAT TGCAGCACTC
      2260        2270        2280        2290        2300
CGCGAGTCTA TCCAACAGAC ACTTGTCGAG TCCTCTCAAT GGGTAGCTGG
      2310        2320        2330        2340        2350
TGAGCAACAA GTCAAGTGGA AACGCCGCTG GATTAAAGGT CTGGATGGTC
      2360        2370        2380        2390        2400
GTAAGGTACA CGTTCGTAGT CCTCACGCTG CCTTGAATAC CCTACTGCAA
      2410        2420        2430        2440        2450
TCTGCTGGTG CTCTCATCTG CAAACTGTGG ATTATCAAGA CCGAAGAGAT
      2460        2470        2480        2490        2500
GCTCGTAGAG AAAGGCTTGA AGCATGGCTG GGATGGGGAC TTTGCGTACA
      2510        2520        2530        2540        2550
TGGCATGGGT ACATGATGAA ATCCAAGTAG GCTGCCGTAC CGAAGAGATT
      2560        2570        2580        2590        2600
GCTCAGGTGG TCATTGAGAC CGCACAAGAA GCGATGCGCT GGGTTGGAGA
      2610        2620        2630        2640        2650
CCACTGGAAC TTCCGGTGTC TTCTGGATAC CGAAGGTAAG ATGGGTCCTA
      2660        2670        2680        2690        2700
ATTGGGCGAT TTGCCACTGA TACAGGAGGC TACTCATGAA CGAAAGACAC
      2710        2720        2730        2740        2750
TTAACAGGTG CTGCTTCTGA AATGCTAGTA GCCTACAAAT TTACCAAAGC
      2760        2770        2780        2790        2800
TGGGTACACT GTCTATTACC CTATGCTGAC TCAGAGTAAA GAGGACTTGG
      2810        2820        2830        2840        2850
TTGTATGTAA GGATGGTAAA TTTAGTAAGG TTCAGGTTAA AACAGCCACA
      2860        2870        2880        2890        2900
ACGGTTCAAA CCAACACAGG AGATGCCAAG CAGGTTAGGC TAGGTGGATG
      2910        2920        2930        2940        2950
CGGTAGGTCC GAATATAAGG ATGGAGACTT TGACATTCTT GCGGTTGTGG
      2960        2970        2980        2990        3000
TTGACGAAGA TGTGCTTATT TTCACATGGG ACGAAGTAAA AGGTAAGACA
      3010        3020        3030        3040        3050
TCCATGTGTG TCGGCAAGAG AAACAAAGGC ATAAAACTAT AGGAGAAATT
      3060        3070        3080
ATTATGGCTA TGACAAAGAA ATTTCCGGAT C
```

FIGURE 9

```
        10         20         30         40         50
AATGCTACTA CTATTAGTAG AATTGATGCC ACCTTTTCAG CTCGCGCCCC
        60         70         80         90        100
AAATGAAAAT ATAGCTAAAC AGGTTATTGA CCATTTGCGA AATGTATCTA
       110        120        130        140        150
ATGGTCAAAC TAAATCTACT CGTTCGCAGA ATTGGGAATC AACTGTTACA
       160        170        180        190        200
TGGAATGAAA CTTCCAGACA CCGTACTTTA GTTGCATATT TAAAACATGT
       210        220        230        240        250
TGAGCTACAG CACCAGATTC AGCAATTAAG CTCTAAGCCA TCCGCAAAAA
       260        270        280        290        300
TGACCTCTTA TCAAAAGGAG CAATTAAAGG TACTCTCTAA TCCTGACCTG
       310        320        330        340        350
TTGGAGTTTG CTTCCGGTCT GGTTCGCTTT GAAGCTCGAA TTAAAACGCG
       360        370        380        390        400
ATATTTGAAG TCTTTCGGGC TTCCTCTTAA TCTTTTTGAT GCAATCCGCT
       410        420        430        440        450
TTGCTTCTGA CTATAATAGT CAGGGTAAAG ACCTGATTTT TGATTTATGG
       460        470        480        490        500
TCATTCTCGT TTTCTGAACT GTTTAAAGCA TTTGAGGGGG ATTCAATGAA
       510        520        530        540        550
TATTTATGAC GATTCCGCAG TATTGGACGC TATCCAGTCT AAACATTTTA
       560        570        580        590        600
CTATTACCCC CTCTGGCAAA ACTTCTTTTG CAAAAGCCTC TCGCTATTTT
       610        620        630        640        650
GGTTTTTATC GTCGTCTGGT AAACGAGGGT TATGATAGTG TTGCTCTTAC
       660        670        680        690        700
TATGCCTCGT AATTCCTTTT GGCGTTATGT ATCTGCATTA GTTGAATGTG
       710        720        730        740        750
GTATTCCTAA ATCTCAACTG ATGAATCTTT CTACCTGTAA TAATGTTGTT
       760        770        780        790        800
CCGTTAGTTC GTTTTATTAA CGTAGATTTT TCTTCCCAAC GTCCTGACTG
       810        820        830        840        850
GTATAATGAG CCAGTTCTTA AAATCGCATA AGGTAATTCA CAATGATTAA
       860        870        880        890        900
```

FIGURE 9 (continued)

```
AGTTGAAATT AAACCATCTC AAGCCCAATT TACTACTCGT TCTGGTGGTT
       910        920        930        940        950
CTCGTCAGGG CAAGCCTTAT TCACTGAATG AGCAGCTTTG TTACGTTGAT
       960        970        980        990       1000
TTGGGTAATG AATATCCGGT TCTTGTCAAG ATTACTCTTG ATGAAGGTCA
      1010       1020       1030       1040       1050
GCCAGCCTAT GCGCCTGGTC TGTACACCGT TCATCTGTCC TCTTTCAAAG
      1060       1070       1080       1090       1100
TTGGTCAGTT CGGTTCCCTT ATGATTGACC GTCTGCGCCT CGTTCCGGCT
      1110       1120       1130       1140       1150
AAGTAACATG GAGCAGGTCG CGGATTTCGA CACAATTTAT CAGGCGATGA
      1160       1170       1180       1190       1200
TACAAATCTC CGTTGTACTT TGTTTCGCGC TTGGTATAAT CGCTGGGGGT
      1210       1220       1230       1240       1250
CAAAGATGAG TGTTTTAGTG TATTCTTTCG CCTCTTTCGT TTTAGGTTGG
      1260       1270       1280       1290       1300
TGCCTTCGTA GTGGCATTAC GTATTTTACC CGTTTAATGC AAACTTCCTC
      1310       1320       1330       1340       1350
ATGAAAAAGT CTTTAGTCCT CAAAGCCTCT GTAGCCGTTG CTACCCTCGT
      1360       1370       1380       1390       1400
TCCGATGCTG TCTTTCGCTG CTGAGGGTGA CGATCCCGCA AAAGCGGCCT
      1410       1420       1430       1440       1450
TTAACTCCCT GCAAGCCTCA GCGACCGAAT ATATCGGTTA TGCGTGGGCG
      1460       1470       1480       1490       1500
ATGGTTGTTG TCATTGTCGG CGCAACTATC GGTATCAAGC TGTTTAAGAA
      1510       1520       1530       1540       1550
ATTCACCTCG AAAGCAAGCT GATAAACCGA TACAATTAAA GGCTCCTTTT
      1560       1570       1580       1590       1600
GGAGCCTTTT TTTTTGGAGA TTTTCAACGT GAAAAAATTA TTATTCGCAA
      1610       1620       1630       1640       1650
TTCCTTTAGT TGTTCCTTTC TATTCTCACT CCGCTGAAAC TGTTGAAAGT
      1660       1670       1680       1690       1700
TGTTTAGCAA AACCCCATAC AGAAAATTCA TTTACTAACG TCTGGAAAGA
      1710       1720       1730       1740       1750
CGACAAAACT TTAGATCGTT ACGCTAACTA TGAGGGTTGT CTGTGGAATG
      1760       1770       1780       1790       1800
CTACAGGCGT TGTAGTTTGT ACTGGTGACG AAACTCAGTG TTACGGTACA
      1810       1820       1830       1840       1850
TGGGTTCCTA TTGGGCTTGC TATCCCTGAA AATGAGGGTG GTGGCTCTGA
      1860       1870       1880       1890       1900
GGGTGGCGGT TCTGAGGGTG GCGGTTCTGA GGGTGGCGGT ACTAAACCTC
      1910       1920       1930       1940       1950
CTGAGTACGG TGATACACCT ATTCCGGGCT ATACTTATAT CAACCCTCTC
      1960       1970       1980       1990       2000
GACGGCACTT ATCCGCCTGG TACTGAGCAA AACCCCGCTA ATCCTAATCC
      2010       2020       2030       2040       2050
TTCTCTTGAG GAGTCTCAGC CTCTTAATAC TTTCATGTTT CAGAATAATA
      2060       2070       2080       2090       2100
GGTTCCGAAA TAGGCAGGGG GCATTAACTG TTTATACGGG CACTGTTACT
      2110       2120       2130       2140       2150
CAAGGCACTG ACCCCGTTAA AACTTATTAC CAGTACACTC CTGTATCATC
      2160       2170       2180       2190       2200
AAAAGCCATG TATGACGCTT ACTGGAACGG TAAATTCAGA GACTGCGCTT
      2210       2220       2230       2240       2250
```

FIGURE 9 (continued)

```
TCCATTCTGG CTTTAATGAA GATCCATTCG TTTGTGAATA TCAAGGCCAA
      2260       2270       2280       2290       2300
TCGTCTGACC TGCCTCAACC TCCTGTCAAT GCTGGCGGCG GCTCTGGTGG
      2310       2320       2330       2340       2350
TGGTTCTGGT GGCGGCTCTG AGGGTGGTGG CTCTGAGGGT GGCGGTTCTG
      2360       2370       2380       2390       2400
AGGGTGGCGG CTCTGAGGGA GGCGGTTCCG GTGGTGGCTC TGGTTCCGGT
      2410       2420       2430       2440       2450
GATTTTGATT ATGAAAAGAT GGCAAACGCT AATAAGGGGG CTATGACCGA
      2460       2470       2480       2490       2500
AAATGCCGAT GAAAACGCGC TACAGTCTGA CGCTAAAGGC AAACTTGATT
      2510       2520       2530       2540       2550
CTGTCGCTAC TGATTACGGT GCTGCTATCG ATGGTTTCAT TGGTGACGTT
      2560       2570       2580       2590       2600
TCCGGCCTTG CTAATGGTAA TGGTGCTACT GGTGATTTTG CTGGCTCTTA
      2610       2620       2630       2640       2650
TTCCCAAATG GCTCAAGTCG GTGACGGTGA TAATTCACCT TTAATGAATA
      2660       2670       2680       2690       2700
ATTTCCGTCA ATATTTACCT TCCCTCCCTC AATCGGTTGA ATGTCGCCCT
      2710       2720       2730       2740       2750
TTTGTCTTTA GCGCTGGTAA ACCATATGAA TTTTCTATTG ATTGTGACAA
      2760       2770       2780       2790       2800
AATAAACTTA TTCCGTGGTG TCTTTGCGTT TCTTTTATAT GTTGCCACCT
      2810       2820       2830       2840       2850
TTATGTATGT ATTTTCTACG TTTGCTAACA TACTGCGTAA TAAGGAGTCT
      2860       2870       2880       2890       2900
TAATCATGCC AGTTCTTTTG GGTATTCCGT TATTATTGCG TTTCCTCGGT
      2910       2920       2930       2940       2950
TTCCTTCTGG TAACTTTGTT CGGCTATCTG CTTACTTTTC TTAAAAAGGG
      2960       2970       2980       2990       3000
CTTCGGTAAG ATAGCTATTG CTATTTCATT GTTTCTTGCT CTTATTATTG
      3010       3020       3030       3040       3050
GGCTTAACTC AATTCTTGTG GGTTATCTCT CTGATATTAG CGCTCAATTA
      3060       3070       3080       3090       3100
CCCTCTGACT TTGTTCAGGG TGTTCAGTTA ATTCTCCCGT CTAATGCGCT
      3110       3120       3130       3140       3150
TCCCTGTTTT TATGTTATTC TCTCTGTAAA GGCTGCTATT TTCATTTTTG
      3160       3170       3180       3190       3200
ACGTTAAACA AAAAATCGTT TCTTATTTGG ATTGGGATAA ATAATATGGC
      3210       3220       3230       3240       3250
TGTTTATTTT GTAACTGGCA AATTAGGCTC TGGAAAGACG CTCGTTAGCG
      3260       3270       3280       3290       3300
TTGGTAAGAT TCAGGATAAA ATTGTAGCTG GGTGCAAAAT AGCAACTAAT
      3310       3320       3330       3340       3350
CTTGATTTAA GGCTTCAAAA CCTCCCGCAA GTCGGGAGGT TCGCTAAAAC
      3360       3370       3380       3390       3400
GCCTCGCGTT CTTAGAATAC CGGATAAGCC TTCTATATCT GATTTGCTTG
      3410       3420       3430       3440       3450
CTATTGGGCG CGGTAATGAT TCCTACGATG AAAATAAAAA CGGCTTGCTT
      3460       3470       3480       3490       3500
GTTCTCGATG AGTGCGGTAC TTGGTTTAAT ACCCGTTCTT GGAATGATAA
      3510       3520       3530       3540       3550
GGAAAGACAG CCGATTATTG ATTGGTTTCT ACATGCTCGT AAATTAGTAT
      3560       3570       3580       3590       3600
```

FIGURE 9 (continued)

```
GGGATATTAT TTTTCTTGTT CAGGACTTAT CTATTGTTGA TAAACAGGCG
      3610       3620       3630       3640       3650
CGTTCTGCAT TAGCTGAACA TGTTGTTTAT TGTCGTCGTC TGGACAGAAT
      3660       3670       3680       3690       3700
TACTTTACCT TTTGTCGGTA CTTTATATTC TCTTATTACT GGCTCGAAAA
      3710       3720       3730       3740       3750
TGCCTCTGCC TAAATTACAT GTTGGCGTTG TTAAATATGG CGATTCTCAA
      3760       3770       3780       3790       3800
TTAAGCCCTA CTGTTGAGCG TTGGCTTTAT ACTGGTAAGA ATTTGTATAA
      3810       3820       3830       3840       3850
CGCATATGAT ACTAAACAGG CTTTTTCTAG TAATTATGAT TCCGGTGTTT
      3860       3870       3880       3890       3900
ATTCTTATTT AACGCCTTAT TTATCACACG GTCGGTATTT CAAACCATTA
      3910       3920       3930       3940       3950
AATTTAGGTC AGAAGATGAA ATTAACTAAA ATATATTTGA AAAAGTTTTC
      3960       3970       3980       3990       4000
TCGCGTTCTT TGTCTTGCGA TTGGATTTGC ATCAGCATTT ACATATAGTT
      4010       4020       4030       4040       4050
ATATAACCCA ACCTAAGCCG GAGGTTAAAA AGGTAGTCTC TCAGACCTAT
      4060       4070       4080       4090       4100
GATTTTGATA AATTCACTAT TGACTCTTCT CAGCGTCTTA ATCTAAGCTA
      4110       4120       4130       4140       4150
TCGCTATGTT TTCAAGGATT CTAAGGGAAA ATTAATTAAT AGCGACGATT
      4160       4170       4180       4190       4200
TACAGAAGCA AGGTTATTCA CTCACATATA TTGATTTATG TACTGTTTCC
      4210       4220       4230       4240       4250
ATTAAAAAAG GTAATTCAAA TGAAATTGTT AAATGTAATT AATTTTGTTT
      4260       4270       4280       4290       4300
TCTTGATGTT TGTTTCATCA TCTTCTTTTG CTCAGGTAAT TGAAATGAAT
      4310       4320       4330       4340       4350
AATTCGCCTC TGCGCGATTT TGTAACTTGG TATTCAAAGC AATCAGGCGA
      4360       4370       4380       4390       4400
ATCCGTTATT GTTTCTCCCG ATGTAAAAGG TACTGTTACT GTATATTCAT
      4410       4420       4430       4440       4450
CTGACGTTAA ACTTGAAAAT CTACGCAATT TCTTTATTTC TGTTTTACGT
      4460       4470       4480       4490       4500
GCTAATAATT TTGATATGGT TGGTTCAATT CCTTCCATAA TTCAGAAGTA
      4510       4520       4530       4540       4550
TAATCCAAAC AATCAGGTAT ATATTGATGA ATTGCCATCA TCTGATAATC
      4560       4570       4580       4590       4600
AGGAATATGA TGATAATTCC GCTCCTTCTG GTGGTTTCTT TGTTCCGCAA
      4610       4620       4630       4640       4650
AATGATAATG TTACTCAAAC TTTTAAAATT AATAACGTTC GGGCAAAGGA
      4660       4670       4680       4690       4700
TTTAATACGA GTTGTCGAAT TGTTTGTAAA GTCTAATACT TCTAAATCCT
      4710       4720       4730       4740       4750
CAAATGTATT ATCTATTGAC GGCTCTAATC TATTAGTTGT TAGTGCACCT
      4760       4770       4780       4790       4800
AAAGATATTT TAGATAACCT TCCTCAATTC CTTTCTACTG TTGATTTGCC
      4810       4820       4830       4840       4850
AACTGACCAG ATATTGATTG AGGGTTTGAT ATTTGAGGTT CAGCAAGGTG
      4860       4870       4880       4890       4900
ATGCTTTAGA TTTTTCATTT GCTGCTGGCT CTCAGCGTGG CACTGTTGCA
      4910       4920       4930       4940       4950
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FIGURE 9 (continued)

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GGCGGTGTTA ATACTGACCG CCTCACCTCT GTTTTATCTT CTGCTGGTGG
      4960       4970       4980       4990       5000
TTCGTTCGGT ATTTTTAATG GCGATGTTTT AGGGCTATCA GTTCGCGCAT
      5010       5020       5030       5040       5050
TAAAGACTAA TAGCCATTCA AAAATATTGT CTGTGCCACG TATTCTTACG
      5060       5070       5080       5090       5100
CTTTCAGGTC AGAAGGGTTC TATCTCTGTT GGCCAGAATG TCCCTTTTAT
      5110       5120       5130       5140       5150
TACTGGTCGT GTGACTGGTG AATCTGCCAA TGTAAATAAT CCATTTCAGA
      5160       5170       5180       5190       5200
CGATTGAGCG TCAAAATGTA GGTATTTCCA TGAGCGTTTT TCCTGTTGCA
      5210       5220       5230       5240       5250
ATGGCTGGCG GTAATATTGT TCTGGATATT ACCAGCAAGG CCGATAGTTT
      5260       5270       5280       5290       5300
GAGTTCTTCT ACTCAGGCAA GTGATGTTAT TACTAATCAA AGAAGTATTG
      5310       5320       5330       5340       5350
CTACAACGGT TAATTTGCGT GATGGACAGA CTCTTTTACT CGGTGGCCTC
      5360       5370       5380       5390       5400
ACTGATTATA AAAACACTTC TCAAGATTCT GGCGTACCGT TCCTGTCTAA
      5410       5420       5430       5440       5450
AATCCCTTTA ATCGGCCTCC TGTTTAGCTC CCGCTCTGAT TCCAACGAGG
      5460       5470       5480       5490       5500
AAAGCACGTT ATACGTGCTC GTCAAAGCAA CCATAGTACG CGCCCTGTAG
      5510       5520       5530       5540       5550
CGGCGCATTA AGCGCGGCGG GTGTGGTGGT TACGCGCAGC GTGACCGCTA
      5560       5570       5580       5590       5600
CACTTGCCAG CGCCCTAGCG CCCGCTCCTT TCGCTTTCTT CCCTTCCTTT
      5610       5620       5630       5640       5650
CTCGCCACGT TCGCCGGCTT TCCCCGTCAA GCTCTAAATC GGGGGCTCCC
      5660       5670       5680       5690       5700
TTTAGGGTTC CGATTTAGTG CTTTACGGCA CCTCGACCCC AAAAAACTTG
      5710       5720       5730       5740       5750
ATTTGGGTGA TGGTTCACGT AGTGGGCCAT CGCCCTGATA GACGGTTTTT
      5760       5770       5780       5790       5800
CGCCCTTTGA CGTTGGAGTC CACGTTCTTT AATAGTGGAC TCTTGTTCCA
      5810       5820       5830       5840       5850
AACTGGAACA ACACTCAACC CTATCTCGGG CTATTCTTTT GATTTATAAG
      5860       5870       5880       5890       5900
GGATTTTGCC GATTTCGGAA CCACCATCAA ACAGGATTTT CGCCTGCTGG
      5910       5920       5930       5940       5950
GGCAAACCAG CGTGGACCGC TTGCTGCAAC TCTCTCAGGG CCAGGCGGTG
      5960       5970       5980       5990       6000
AAGGGCAATC AGCTGTTGCC CGTCTCGCTG GTGAAAAGAA AAACCACCCT
      6010       6020       6030       6040       6050
GGCGCCCAAT ACGCAAACCG CCTCTCCCCG CGCGTTGGCC GATTCATTAA
      6060       6070       6080       6090       6100
TCCAGCTGGC ACGACAGGTT TCCCGACTGG AAAGCGGGCA GTGAGCGCAA
      6110       6120       6130       6140       6150
CGCAATTAAT GTGAGTTACC TCACTCATTA GGCACCCCAG GCTTTACACT
      6160       6170       6180       6190       6200
TTATGCTTCC GGCTCGTATG TTGTGTGGAA TTGTGAGCGG ATAACAATTT
      6210       6220       6230       6240       6250
CACACAGGAA ACAGCTATGA CCATGATTAC GAATTCGAGC TCGCCCGGGG
      6260       6270       6280       6290       6300
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FIGURE 9 (continued)

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ATCTGCCTGA ATAGGTACGA TTTACTAACT GGAAGAGGCA CTAAATGAAC
      6310       6320       6330       6340       6350
ACGATTAACA TCGCTAAGAA CGACTTCTCT GACATCGAAC TGGCTGCTAT
      6360       6370       6380       6390       6400
CCCGTTCAAC ACTCTGGCTG ACCATTACGG TGAGCGTTTA GCTCGCGAAC
      6410       6420       6430       6440       6450
AGTTGGCCCT TGAGCATGAG TCTTACGAGA TGGGTGAAGC ACGCTTCCGC
      6460       6470       6480       6490       6500
AAGATGTTTG AGCGTCAACT TAAAGCTGGT GAGGTTGCGG ATAACGCTGC
      6510       6520       6530       6540       6550
CGCCAAGCCT CTCATCACTA CCCTACTCCC TAAGATGATT GCACGCATCA
      6560       6570       6580       6590       6600
ACGACTGGTT TGAGGAAGTG AAAGCTAAGC GCGGCAAGCG CCCGACAGCC
      6610       6620       6630       6640       6650
TTCCAGTTCC TGCAAGAAAT CAAGCCGGAA GCCGTAGCGT ACATCACCAT
      6660       6670       6680       6690       6700
TAAGACCACT CTGGCTTGCC TAACCAGTGC TGACAATACA ACCGTTCAGG
      6710       6720       6730       6740       6750
CTGTAGCAAG CGCAATCGGT CGGGCCATTG AGGACGAGGC TCGCTTCGGT
      6760       6770       6780       6790       6800
CGTATCCGTG ACCTTGAAGC TAAGCACTTC AAGAAAAACG TTGAGGAACA
      6810       6820       6830       6840       6850
ACTCAACAAG CGCGTAGGGC ACGTCTACAA GAAAGCATTT ATGCAAGTTG
      6860       6870       6880       6890       6900
TCGAGGCTGA CATGCTCTCT AAGGGTCTAC TCGGTGGCGA GGCGTGGTCT
      6910       6920       6930       6940       6950
TCGTGGCATA AGGAAGACTC TATTCATGTA GGAGTACGCT GCATCGAGAT
      6960       6970       6980       6990       7000
GCTCATTGAG TCAACCGGAA TGGTTAGCTT ACACCGCCAA AATGCTGGCG
      7010       7020       7030       7040       7050
TAGTAGGTCA AGACTCTGAG ACTATCGAAC TCGCACCTGA ATACGCTGAG
      7060       7070       7080       7090       7100
GCTATCGCAA CCCGTGCAGG TGCGCTGGCT GGCATCTCTC CGATGTTCCA
      7110       7120       7130       7140       7150
ACCTTGCGTA GTTCCTCCTA AGCCGTGGAC TGGCATTACT GGTGGTGGCT
      7160       7170       7180       7190       7200
ATTGGGCTAA CGGTCGTCGT CCTCTGGCGC TGGTGCGTAC TCACAGTAAG
      7210       7220       7230       7240       7250
AAAGCACTGA TGCGCTACGA AGACGTTTAC ATGCCTGAGG TGTACAAAGC
      7260       7270       7280       7290       7300
GATTAACATT GCGCAAAACA CCGCATGGAA AATCAACAAG AAAGTCCTAG
      7310       7320       7330       7340       7350
CGGTCGCCAA CGTAATCACC AAGTGGAAGC ATTGTCCGGT CGAGGACATC
      7360       7370       7380       7390       7400
CCTGCGATTG AGCGTGAAGA ACTCCCGATG AAACCGGAAG ACATCGACAT
      7410       7420       7430       7440       7450
GAATCCTGAG GCTCTCACCG CGTGGAAACG TGCTGCCGCT GCTGTGTACC
      7460       7470       7480       7490       7500
GCAAGGACAA GGCTCGCAAG TCTCGCCGTA TCAGCCTTGA GTTCATGCTT
      7510       7520       7530       7540       7550
GAGCAAGCCA ATAAGTTTGC TAACCATAAG GCCATCTGGT TCCCTTACAA
      7560       7570       7580       7590       7600
CATGGACTGG CGCGGTCGTG TTTACGCTGT GTCAATGTTC AACCCGCAAG
      7610       7620       7630       7640       7650
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FIGURE 9 (continued)

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GTAACGATAT GACCAAAGGA CTGCTTACGC TGGCGAAAGG TAAACCAATC
      7660       7670       7680       7690       7700
GGTAAGGAAG GTTACTACTG GCTGAAAATC CACGGTGCAA ACTGTGCGGG
      7710       7720       7730       7740       7750
TGTCGATAAG GTTCCGTTCC CTGAGCGCAT CAAGTTCATT GAGGAAAACC
      7760       7770       7780       7790       7800
ACGAGAACAT CATGGCTTGC GCTAAGTCTC CACTGGAGAA CACTTGGTGG
      7810       7820       7830       7840       7850
GCTGAGCAAG ATTCTCCGTT CTGCTTCCTT GCGTTCTGCT TTGAGTACGC
      7860       7870       7880       7890       7900
TGGGGTACAG CACCACGGCC TGAGCTATAA CTGCTCCCTT CCGCTGGCGT
      7910       7920       7930       7940       7950
TTGACGGGTC TTGCTCTGGC ATCCAGCACT TCTCCGCGAT GCTCCGAGAT
      7960       7970       7980       7990       8000
GAGGTAGGTG GTCGCGCGGT TAACTTGCTT CCTAGTGAAA CCGTTCAGGA
      8010       8020       8030       8040       8050
CATCTACGGG ATTGTTGCTA AGAAAGTCAA CGAGATTCTA CAAGCAGACG
      8060       8070       8080       8090       8100
CAATCAATGG GACCGATAAC GAAGTAGTTA CCGTGACCGA TGAGAACACT
      8110       8120       8130       8140       8150
GGTGAAATCT CTGAGAAAGT CAAGCTGGGC ACTAAGGCAC TGGCTGGTCA
      8160       8170       8180       8190       8200
ATGGCTGGCT TACGGTGTTA CTCGCAGTGT GACTAAGCGT TCAGTCATGA
      8210       8220       8230       8240       8250
CGCTGGCTTA CGGGTCCAAA GAGTTCGGCT TCCGTCAACA AGTGCTGGAA
      8260       8270       8280       8290       8300
GATACCATTC AGCCAGCTAT TGATTCCGGC AAGGGTCTGA TGTTCACTCA
      8310       8320       8330       8340       8350
GCCGAATCAG GCTGCTGGAT ACATGGCTAA GCTGATTTGG GAATCTGTGA
      8360       8370       8380       8390       8400
GCGTGACGGT GGTAGCTGCG GTTGAAGCAA TGAACTGGCT TAAGTCTGCT
      8410       8420       8430       8440       8450
GCTAAGCTGC TGGCTGCTGA GGTCAAAGAT AAGAAGACTG GAGAGATTCT
      8460       8470       8480       8490       8500
TCGCAAGCGT TGCGCTGTGC ATTGGGTAAC TCCTGATGGT TTCCCTGTGT
      8510       8520       8530       8540       8550
GGCAGGAATA CAAGAAGCCT ATTCAGACGC GCTTGAACCT GATGTTCCTC
      8560       8570       8580       8590       8600
GGTCAGTTCC GCTTACAGCC TACCATTAAC ACCAACAAAG ATAGCGAGAT
      8610       8620       8630       8640       8650
TGATGCACAC AAACAGGAGT CTGGTATCGC TCCTAACTTT GTACACAGCC
      8660       8670       8680       8690       8700
AAGACGGTAG CCACCTTCGT AAGACTGTAG TGTGGGCACA CGAGAAGTAC
      8710       8720       8730       8740       8750
GGAATCGAAT CTTTTGCACT GATTCACGAC TCCTTCGGTA CCATTCCGGC
      8760       8770       8780       8790       8800
TGACGCTGCG AACCTGTTCA AAGCAGTGCG CGAAACTATG GTTGACACAT
      8810       8820       8830       8840       8850
ATGAGTCTTG TGATGTACTG GCTGATTTCT ACGACCAGTT CGCTGACCAG
      8860       8870       8880       8890       8900
TTGCACGAGT CTCAATTGGA CAAAATGCCA GCACTTCCGG CTAAAGGTAA
      8910       8920       8930       8940       8950
CTTGAACCTC CGTGACATCT TAGAGTCGGA CTTCGCGTTC GCGTAACGCC
      8960       8970       8980       8990       9000
```

FIGURE 9 (continued)

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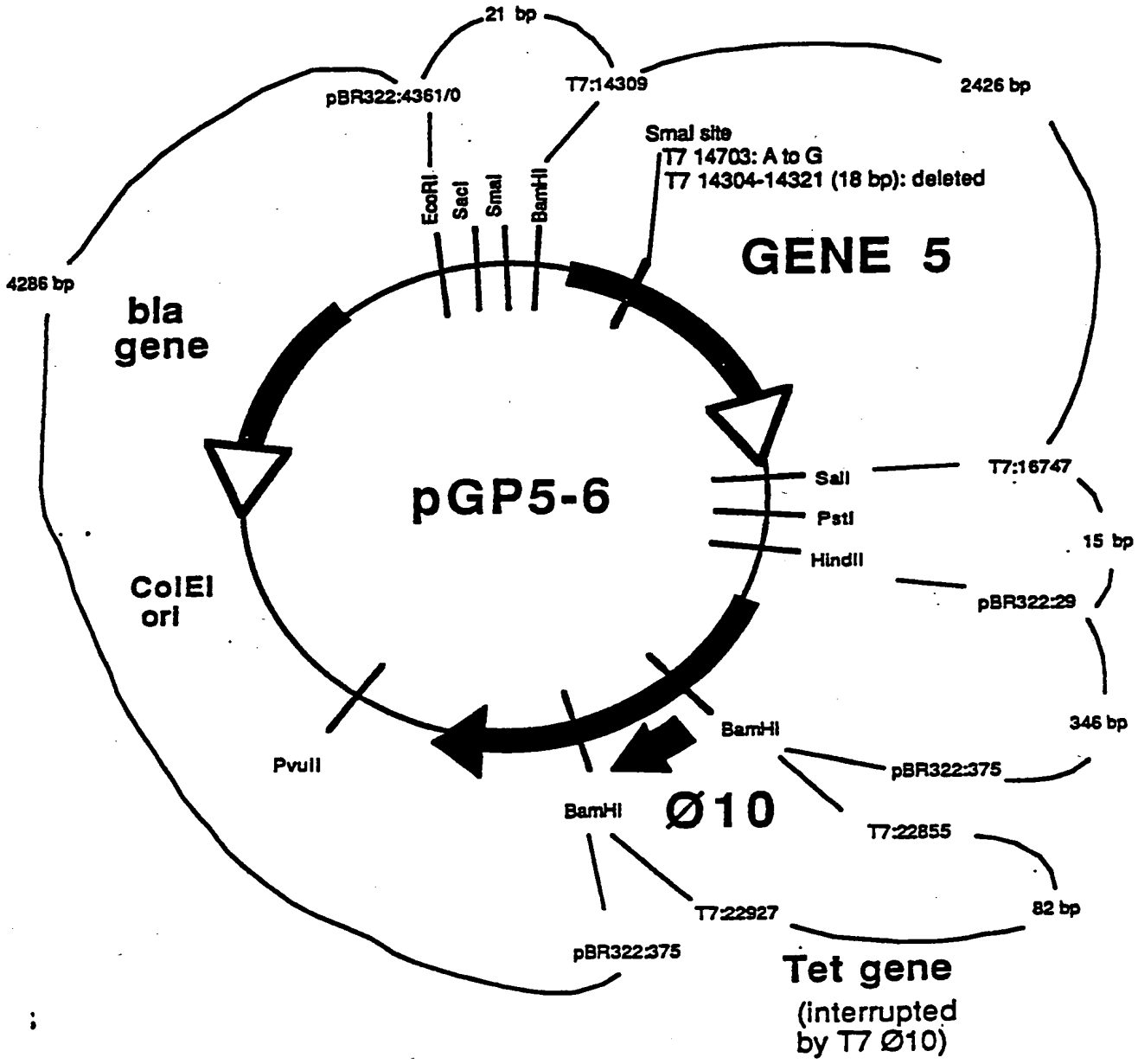
AAATCAATAC GACCCGGATC GGTCGACCTG CAGCCCAAGC TTGGCACTGG
      9010        9020        9030        9040        9050
CCGTCGTTTT ACAACGTCGT GACTGGGAAA ACCCTGGCGT TACCCAACTT
      9060        9070        9080        9090        9100
AATCGCCTTG CAGCACATCC CCCCTTCGCC AGCTGGCGTA ATAGCGAAGA
      9110        9120        9130        9140        9150
GGCCCGCACC GATCGCCCTT CCCAACAGTT GCGTAGCCTG AATGGCGAAT
      9160        9170        9180        9190        9200
GGCGCTTTGC CTGGTTTCCG GCACCAGAAG CGGTGCCGGA AAGCTGGCTG
      9210        9220        9230        9240        9250
GAGTGCGATC TTCCTGAGGC CGAQACNGTC GTCGTCCCCT CAAACTGGCA
      9260        9270        9280        9290        9300
GATGCACGGT TACGATGCGC CCATCTACAC CAACGTAACC TATCCCATTA
      9310        9320        9330        9340        9350
CGGTCAATCC GCCGTTTGTT CCCACGGAGA ATCCGACGGG TTGTTACTCG
      9360        9370        9380        9390        9400
CTCACATTTA ATGTTGATGA AAGCTGGCTA CAGGAAGGCC AGACGCGAAT
      9410        9420        9430        9440        9450
TATTTTTGAT GGCGTTCCTA TTGGTTAAAA AATGAGCTGA TTTAACAAAA
      9460        9470        9480        9490        9500
ATTTAACGCG AATTTTAACA AAATATTAAC GTTTACAATT TAAATATTTG
      9510        9520        9530        9540        9550
CTTATACAAT CTTCCTGTTT TTGGGGCTTT TCTGATTATC AACCGGGGTA
      9560        9570        9580        9590        9600
CATATGATTG ACATGCTAGT TTTACGATTA CCGTTCATCG ATTCTCTTGT
      9610        9620        9630        9640        9650
TTGCTCCAGA CTCTCAGGCA ATGACCTGAT AGCCTTTGTA GATCTCTCAA
      9660        9670        9680        9690        9700
AAATAGCTAC CCTCTCCGGC ATGAATTTAT CAGCTAGAAC GGTTGAATAT
      9710        9720        9730        9740        9750
CATATTGATG GTGATTTGAC TGTCTCCGGC CTTTCTCACC CTTTTGAATC
      9760        9770        9780        9790        9800
TTTACCTACA CATTACTCAG GCATTGCATT TAAAATATAT GAGGGTTCTA
      9810        9820        9830        9840        9850
AAAATTTTTA TCCTTGCGTT GAAATAAAGG CTTCTCCCGC AAAAGTATTA
      9860        9870        9880        9890        9900
CAGGGTCATA ATGTTTTTGG TACAACCGAT TTAGCTTTAT GCTCTGAGGC
      9910        9920        9930        9940        9950
TTTATTGCTT AATTTTGCTA ATTCTTTGCC TTGCCTGTAT GATTTATTGG

ATGTT
```

FIGURE 10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2426 bp
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SacI
SmaI
BamHI
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gene
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SalI
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